Long QT Syndrome

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SUMMARY

Long QT syndrome (LQTS) is a heart rhythm disorder characterized by QT prolongation and T wave abnormalities associated with increased susceptibility to torsade de pointes (TdP) that can lead to cardiac events (CE: syncope, cardiac arrest and sudden death). The prevalence of congenital LQTS is estimated as 1:2500 caused by mutations of genes encoding or regulating cardiac sodium, potassium, and calcium ion channels. To date 14 subtypes including 12 in Romano-Ward syndrome and 2 in Jervell and Lange-Nielsen syndrome caused by 12 genes with >1200 mutations have been identified. Most of the known mutations encode potassium ion channels. The disease severity is influenced by the mutation type, location and ion channel biophysical properties. Clinically LQTS-related symptoms are related to age, gender and the degree of QT prolongation. The CE rate is the highest among teenagers and young adults. Females are more problematic than males. The longer QT interval, the higher is the CE rate. Among genotyped individuals LQT1-3 account for 90-95%. Gene-specific ECG phenotype is present in LQT1-3 and 7. The triggers to CEs also differ by genotype. Recognizing gene-specific clinical features and ECG patterns can improve diagnostic accuracy. Family screening, serial ECG follow-up, and stress testing are important not only for the diagnosis of probands, but also for identifying additional affected family members. As the mainstay therapy to all subtypes, β -blocker medication is the most effective to LQT1. Left cardiac sympathetic denervation (LCSD) is known to raise the threshold for ventricular fibrillation and has shown the promise to reduce the CE rate. The minimum invasive approach can provide safer and faster recovery than the traditional LCSD. Implantable cardioverter defibrillators (ICD) provide the best protection against lifethreatening arrhythmias. However, risk stratification for ICD applications is necessary since up to 50% of LQTS never experience CEs in their life time. Acquired LQTS is much more common in the general population and females are more susceptible to QT prolonging drugs and electrolyte imbalance induced TdP. Removing the cause is the best way to redress acquired LQTS. Avoiding QT prolonging drugs and gene-specific triggers can lower the CE rate in congenital LQTS as well.

OUTLINE

From genes to patients, this article provides an overview of current understanding of long QT syndrome (LQTS) in both congenital and acquired forms. In the congenital form, 14 subtypes of LQTS including 12 in Romano-Ward syndrome and 2 in Jervell and Lange-Nielsen syndrome caused by 12 genes encoding cardiac sodium, potassium and calcium ion channel subunits are described. Among them LQT1-3 are the common genotypes that account for 90-95% of gene carriers. The clinical course, findings in genotype-phenotype correlation studies, diagnostic and treatment strategies are discussed. In the acquired form, several common causes especially drug-induced LQTS with the underpinning QT prolonging mechanism are provided. With better understanding of the natural course, improved diagnostic methodology/technology and effective therapeutics, the prognosis is optimistic. LQTS is treatable and sudden death can be prevented.

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Key words >

Long QT syndrome - Sudden cardiac death - Arrhythmias, cardiac

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АКАР	Akinase anchoring protein	NMD	Nonsense-mediated decay
ANKB	Ankyrin B	nNos	Neuronal nitric oxide synthase
APD	Action potential duration	РКА	Kinase A
AT-PASE	Adenosine triphosphatase	PMLA4b	Plasma membrane Ca-ATPhase subtype4b
ATS1	Type 1 Andersen-Tawil syndrome	PP1	Phosphatase 1
CAV3	Caveolin 3	PUCs	Premature ventricular contractions
CE	Cardiac events	RWs	Romano-Ward syndrome
DAD	Delayed-afterdepolarization	SAH	Subarachnoid hemorrhage
EAD	Early-after-depolarization	TdP	Torsade de Pointes
ICD	Implantable cardioverter defibrillator	TDR	Transmural dispersion of repolarization
JLN1	Type1 JLNS	тs	Timothy syndrome
JLN2	Type2 JLNN	TS1	Type-1TS
JLNS	Jervell and Lange-Nielsen syndrome	TS2	Type-2TS
LCSD	left cardiac sympathetic denervation	VF	ventricular fibrillation
LQTS	Long QT syndrome		
	AKAP ANKB APD AT-PASE ATS1 CAV3 CE DAD EAD ICD JLN1 JLN2 JLNS LCSD LQTS	AKAPAkinase anchoring proteinANKBAnkyrin BAPDAction potential durationATPASEAdenosine triphosphataseATS1Type 1 Andersen-Tawil syndromeCAV3Caveolin 3CECardiac eventsDADDelayed-afterdepolarizationEADEarly-after-depolarizationICDImplantable cardioverter defibrillatorJLN1Type2 JLNNJLN2Jervell and Lange-Nielsen syndromeLCSDLong QT syndrome	AKAPAkinase anchoring proteinNMDANKBAnkyrin BnNosAPDAction potential durationPKAAT-PASEAdenosine triphosphatasePMLA4bAT51Type 1 Andersen-Tawil syndromePP1CAV3Caveolin 3PUCsCECardiac eventsRWsDADDelayed-afterdepolarizationSAHICDImplantable cardioverter defibrillatorTDRJLN1Type2 JLNNTS1JLN2Jervell and Lange-Nielsen syndromeVFLQTSLong QT syndromeV

INTRODUCTION

Long QT syndrome (LQTS) is a heart rhythm disorder. (1-4) Patients with LQTS have a propensity to polymorphic ventricular arrhythmias, typically torsade de pointes (TdP). (5-10) In most cases TdP is self-terminating. However, under certain circumstances, TdP can deteriorate into ventricular fibrillation (VF), causing cardiac arrest and ultimately sudden death. (11) Physical and emotional stress can trigger lifethreatening arrhythmias in LQTS. (12-14)

Collectively LQTS is categorized as a channelopathy (15-17) Channelopathies are diseases caused by disturbed function of ion channel subunits or the proteins that regulate them (18) Ion channels have specific ion selectivity and allow the passage of charged ions, such as sodium, potassium and calcium, across the cell membrane. The precision and timeliness of the passage of these charged ions mediated by their specific ion channel proteins provide the molecular biophysical basis for cardiac electrical activity. The P-QRS-T waves are the summations of cardiac ion channel activities registered on the body surface ECG (19-21) Expression of abnormal sodium, calcium, or potassium channels results in aberrant ionic fluxes that can delay ventricular repolarization, manifest as a prolonged QT interval (21) Investigations in LQTS has provided a wealth of information about fundamental mechanisms underlying human cardiac electrophysiology. LQTS can be divided into congenital and acquired forms.

I. Congenital LQTS

Most of congenital LQTS are familial. Very few are due to spontaneous de novo mutations or mosaic mutations. Cardiac events (CE: syncope, cardiac arrest or sudden death) occur most frequently in young otherwise healthy individuals without structural heart abnormalities. The prevalence of LQTS is estimated as 1:2500 (22)

Autosomal dominant inheritance is the most common form, called Romano-Ward syndrome (RWS) and caused by heterozygous mutations in at least 12 different genes (Table 1) (23). Six of the genes encode cardiac potassium channels and most of the known mutations are located in the potassium channel genes. Potassium channels are made up of several protein subunits. Each subunit is produced from a particular gene. Each channel includes four α (alpha) subunits, which are usually identical, and several β (beta) subunits. α subunits form the hole (pore) through which potassium ions can flow (Figs. 1-2). β subunits help regulate the channel function and interact with various proteins inside and outside the cell. (Figs. 2-3) (24) Four genes encode or regulate cardiac sodium channels and one gene encodes a cardiac calcium channel (Figs. 1-3).

Autosomal recessive inheritance is a rare form, associated with congenital deafness (Jervell and Lange-Nielsen syndrome, JLNS). (1, 25) Two subtypes (JLN1 and JLN2) have been identified based on the disease-causing genes that encode one potassium ion channel (Table 1).

Genotype

LQT1 is the most common genotype in RWS, caused by mutations of *KCNQ1* (26) *KCNQ1* encodes α -subunit of a voltage-dependent potassium channel expressed in various cell types including cardiac myocytes and epithelial cells. In the heart, *KCNQ1* protein assembles with *KCNE1* protein to form Kv7.1, a channel complex constituting the slow component of the delayed rectifier current I_{Ks.} (27) The majority of *KCNQ1* mutations are single nucleotide changes causing single amino acid substitutions in the channel protein (missense mutations). (28) The LQT1-causing mutations have dominant-negative loss-of-function properties. (29) Secondly, I_{Ks} is an adrenergic-sensitive potassium current. CE in LQT1 patients are mostly

Table 1. Genetic basis o	of 12 subtypes of inherited LQTS	
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Туре	Prevalence (among genotyped)	Chromosome / Gene	Protein	Protein function	Mutation effect			
Romano-Ward syndrome (autosomal dominant inheritance)								
LQT1	45-50%	11p15.5 / KCNQ1	K _v 7.1	α -subunit of I _{Ks} channel	$I_{ks} \downarrow$			
LQT2	40-45%	7q35-36 / KCNH2	K _v 11.1	α -subunit of I _{kr} channel	$I_{\kappa r} \downarrow$			
LQT3	3-8%	3p21-24 / SCN5A	Na _v 1.5	α -subunit of Na ⁺ channel	I _{Na.L} ↑			
LQT4	<1%	4q25-27 / ANK2	Ankyrin B	Adapter protein	unclear			
LQT5	<1%	21q22.1 / KCNE1	minK	β -subunit of I _{ks} channel	I _{Ks} ↓			
LQT6	<1%	21a22.1 / KCNE2	MiRP1	β -subunit of I _{kr} channel	$I_{\kappa r} \downarrow$			
LQT7	<2%	17q23 / KCNJ2	Kir2.1	α -subunit of I _{K1} channel	I _{K1} ↓			
LQT8	<1%	12p13 / CACNA1C	Ca, 1.2	lpha-subunit of Ca ²⁺ channel	I _{Cal} ↑			
LQT9	<0.5%	3p25 / CAV3	Caveolin-3	co-localizes with Na, 1.5 at sarcolemma	_{Nal} '!			
LQT10	<0.1%	11q23.3 / SCN4B	â-4	β-subunit of Na⁺ channel	′!			
LQT11	<0.1%	7q21-22 / AKAP9	Yotiao	Mediate I_{κ_s} channel phosphorylation	$I_{\kappa_{s}}\downarrow$			
LQT12	<0.1%	20q11.2/ SNTA1	α 1-syntrophin	Regulates Na ⁺ channel function	I _{Nal} ↑			
Jervell, Lange-Nielsen syndrome (autosomal recessive inheritance)								
JLN1	<0.5%	11p15.5 / KCNQ1	K,7.1	α -subunit of I _K channel	$ _{\kappa_{\varsigma}}\downarrow$			
JLN2	<0.5%	21q22.1 / KCNE1	minK	β -subunit of I _{ks} channel	$I_{\rm Ks}\downarrow$			

Fig 1. Structural models of Pore forming (α) subunits of cardiac sodium (Na_{v)} and potassium (K_v) channels linked LQTS

Part A: Depicting four domains (I-IV) of SCN5A linked to LQT3. Part B & C: Depicting subunits linked to LQT2 and LQT1, respectively.

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triggered by physical stress, notably diving or swimming. (13) Exercise aggravates QT prolongation. (30) As anticipated LQT1 patients respond very well to β -blocker treatment. (31) LQT2 is the second common genotype in RWS, caused by the mutations in *KCNH2* or human ethera-go-go related gene (hERG) that encodes α -subunit of Kv11.1 channel specifically for the rapidly activating delayed rectifier K+ current ($I_{\rm Kr}$) in the heart.(32) *KCNH2* mutations cause loss of Kv11.1 channel function. Different LQT2 mutations cause hERG channel dysfunction by different mechanisms and five mechanisms have been classified, including disruption of Kv11.1 channel synthesis (class 1), protein trafficking (class 2), gating (class 3), permeation (class 4), and degradation of PTC-containing mRNA transcripts by nonsense-mediated mRNA decay (class 5). (33) Class 2 mechanism, the trafficking defects (34, 35) causing reduced delivery of channel protein to the cell surface membrane, is the most common mechanism in hERG dysfunctions. (36, 37)

LQT3 is less common than LQT1 and LQT2, caused by mutations of *SCN5A* that encodes the pore forming á-subunit (Figs. 1-3) of Na⁺ channels (Na_v1.5), referred to as I_{Na^*} (38-41) Voltage-gated Na⁺ channels are transmembrane proteins that produce fast inward I_{Na} currents responsible for the depolarization phase of the cardiac action potential. (24, 42) Na⁺ channels play fundamental roles in the initiation, propagation, and maintenance of normal cardiac rhythm. (24, 43, 44) Depolarizing current through the channel late in the action potential is thought to prolong the action potential duration (APD). The late current is due to failure of the channel to remain inactivated and hence enter a bursting mode in which significant current can enter when it should not. (24, 38-42)

Differed by mutant Na_v1.5 channel biophysics, a gain-of-function mechanism is seen in LQT3 (38-41) and sudden infant death syndrome (SIDS).(45-46) Loss-of-function mutations result in Brugada syndrome (47, 48) and cardiac conduction disease. (49-51) It is of interest that both gain-of-function and loss-of-function can cause sinus node dysfunction, (52) atrial standstill, (53) atrial fibrillation (54-56) and dilated cardiomyopathy. (57) The overlap syndromes of cardiac sodium channel diseases reflect a wide spectrum of phenotypes seen in patients with *SCN5A* mutations. (58-60)

LQT4 is rare, caused by ankyrin B (ANKB)mutations. The phenotype is not as unique as seen in LQT1, LQT2, or LQT3 and QT prolongation is not a hallmark of this entity. A more accurate term for LQT4 is ANKB syndrome. (61-65) Ankyrins are adapter proteins (Fig. 3) that bind to several ion channel proteins, such as chloride-bicarbonate exchanger, sodium-potassium adenosine triphosphatase (ATPase), the voltage-sensitive sodium channel, the sodiumcalcium exchanger (NCX, or $\boldsymbol{I}_{_{Na\text{-}Ca}}),$ and calciumrelease channels including those mediated by the receptors for inositol triphosphate [IP3] or ryanodine. Mutations in this gene interfere with several of these ion channels. (62-65) Such complexity results in a wide spectrum of ECG phenotype presentations, mainly sick sinus syndrome, atrial fibrillation, T-U wave abnormalities and exercise induced ventricular



Fig 2. Structural model of cardiac Ca $_{v}$ (and Na $_{v}$), K $_{v}$, and Kir channels

Top: Four domains (I-IV) of Ca_v (Na_v) individual α -subunits forming Ca_v (Na_v) channels, four domains of individual α subunits forming K_v and Kir channels.

Bottom: Depiction of cardiac Na_v, Ca_v, and K_v channels composed of pore forming α -subunits and variety of accessory subunits.

Reproduced with permission from Publisher and Jeanne Nerbonn, PhD., the author of Molecular Physiology of Cardiac Repolarization, Physiol Rev 85: 1205–1253, 2005²⁴ arrhythmia. The majority of these patients do not show a prolonged QT interval.

LQT5 is caused by mutations of the *KCNE1*, a member of *KCNE* gene family, that encodes mink, a â-subunit of $K_v7.1$ that assembles with α -subunit to form $I_{\rm Ks}$ (Figs. 2-3). *KCNE1* is a modifier gene to $K_v7.1$ and Kv11.1 channel functions. *KCNE1* mutations can result in loss-of-function of $I_{\rm Ks}$ and $I_{\rm Kr}$, thereby reduce outward potassium currents causing delayed repolarization. (66-69)

LQT6 is due to mutations of *KCNE2*, the second member of *KCNE* gene family encoding minK related peptide (MiRP) that coassembles with hERG protein (Fig. 2) to form Kv11.1 channel. Like other *KCNE* isoforms *KCNE2* is also a modifier gene. Mutations in MiRP result in loss-of-function of hERG, therefore reduce I_{Kr} and prolong the QT interval. (258, 70-72) *KCNE2* can affect biophysical properties of K_v 7.1 as well.

LQT7 is also referred as type-1 Andersen-Tawil syndrome (ATS1) caused by *KCNJ2* mutations. (73-75) *KCNJ2* encodes α -subunit of Kir2.1 isoform of Kir2X channels (76) (Fig. 2) that conduct inward rectifier potassium currents I_{K1} .(77) Message and protein expression studies indicate that Kir2.1 is the most abundant Kir2X subfamily member in human ventricle. (78) I_{K1} plays a key role in the final phase of repolarization and maintaining the resting membrane potential. (77) *KCNJ2* mutations cause dominant negative effects of Kir2.1 channel function when co-expressed with wild type subunit. (79) The prominent U wave and QU interval prolongation are the signature changes of ATS1. Perhaps long QU syndrome is a more accurate description of LQT7 since the majority of ATS patients

do not show a prolonged QT interval. (80) Patients with ATS1 are usually in short statue and associated dysmorphic features such as low set of ears and a small chin. The dysmorphic facial features constitute a distinctive trait of ATS, indicating Kir2.1 protein may play a major role in developmental signaling. About 50% of patients have periodic paralysis,(80) in which potassium-sensitive is the most common type.

LQT8 is also referred as Timothy syndrome (TS), a rare type of LQTS caused by CACNA1C mutations. CACNA1C encodes α -subunit (Fig. 2) of the voltagegated calcium channel (Cav1.2) that conducts L-type calcium currents $I_{\ensuremath{\scriptscriptstyle Ca-L}}.$ Two subtypes of TS were reported by Splawski, et al. (81, 82) Type-1 TS (TS1) is caused by a missense mutation (G406R) in exon 8a, an alternative splicing form of exon 8. Most children with TS1 were due to the de novo mutation. In one child parental mosaicism was demonstrated. (81) TS1 children presented with multiorgan dysfunction including markedly prolonged ST-segment and QT interval, lethal arrhythmias, syndactyly, congenital heart disease, immune deficiency, intermittent hypoglycemia, cognitive abnormalities, and autism. (81) The average age of death due to complications of multiorgan dysfunction is 2.5 years. In type-2 (TS2) two mutations G406R and G402S in exon 8 of CACNA1C were identified in two unrelated children showing a severe LQTS phenotype but without syndactyly. (82) One child also had severe mental retardation and nemaline rod skeletal myopathy. Functional expression revealed that the G406R mutation slowed deactivation thus prolonged APD (83) G402S reduced channel inactivation that yielded maintained depolarizing L-type calcium currents. (82)

Fig 3. Structural model depicting the protein-protein interactions involved in regulating/modulating the expression, distribution, and function of cardiac ion channels. The α and β subunits of Na. channel interact with actin cytoskeleton via syntrophindystrophin and ankyrin B and with extracellular matrix via sarcoglycan complex. Caveolin-3 directly interacts with neuronal nitric oxide synthase (nNOS). The K channel's α and β subunits interact with actin cytoskeleton via actin binding proteins filamin and α -actinin and through PDZ domain containing scaffolding proteins. Reproduced with permission from Publisher and Jeanne Nerbonn, PhD., the author of Molecular Physiology of Cardiac Repolarization, Physiol Rev 85: 1205-1253, 200524



LQT9 is caused by mutations in caveolin 3 (*CAV3*) gene.(84) CAV3 encodes for the adapter protein (Fig. 3). Functional studies showed that the mutant *CAV3* results in 2-3 fold increase in $I_{Na,L}$ compared with wild type.(84)The gain-of-function mutations in *CAV3* are also associated with SIDS. (85)

LQT10 is caused by mutations in *SCNB4*, a gene encoding one of four α subunits of voltage-gated Na⁺ channel (Figs. 2-3). These α subunits interact with voltage-gated α subunits to change sodium channel kinetics. The encoded transmembrane protein forms inter-chain disulfide bonds with *SCN2A*. Mutation in this gene can cause long QT syndrome. (86)

LQT11 is caused by the mutation of Yotiao, an Akinase anchoring protein (AKAP). (87) It mediates the formation of a macromolecular complex consisting of the I_{Ks} channel, kinase A (PKA), and phosphatase 1 (PP1). Mutations that disrupt this protein complex do not allow the channel to be regulated in response to stress and can cause death. A heterozygous mutation S1570L in the AKAP9 gene is found in one single Caucasian family with LQTS. (87)

LQT12 is due to syntrophin (*SNTA1*) mutation A390V. (88) The mechanism of LQTS is via the activation of neuronal nitric oxide synthase (nNOS)-*SCN5A* macromolecular complex. A390V can disrupt the binding with plasma membrane Ca-ATPase subtype 4b (PMCA4b) and release inhibition of nNOS, causing S-nitrosylation of *SCN5A* that is associated with increased late sodium current, resulting in a characteristic biophysical dysfunction for sodiumchannel seen in LQT3. These results establish a *SNTA1*-based nNOS complex attached to *SCN5A* as a key regulator of sodium current and suggest that *SNTA1* should be considered a LQTS-susceptibility gene. (88)

JLNS and other compound mutation-caused LQTS are rare. Both compound heterozygous and homozygous mutations of *KCNQ1* and *KCNE1* are reported to cause type-1 JLNS (JLN1) and type-2 JLNS (JLN2) that are associated with congenital deafness (89-93) Compound mutations without deafness have also been reported.(94, 95) As anticipated >1 mutations caused LQTS usually present with severe clinical presentations such as markedly prolonged QT interval, and T wave abnormalities associated with a high risk of arrhythmic sudden death. (1, 25)

Most LQTS mutations identified are missence mutations. (28, 96) It is anticipated that more mutations will be identified in the known genes and the list of LQTS genotypes will continue to grow upon new gene discoveries. DNA sequences in the noncoding regions cannot be overlooked because intronic variants can cause LQTS. (97) Moreover, up to 10% of patients have large gene deletions or duplications among major known LQTS susceptibility genes that cannot be detected by sequencing-based mutation screening. (98)

Genotype-phenotype correlation

Among genotyped patients, LQT1, LQT2, and LQT3 account for 90-95% (99) and LQT4 to LQT12 are rare. A series of genotype-phenotype correlation studies have been conducted in regard to mutation types and locations, triggers of cardiac events and ECG characteristics. Recognizing genotype-phenotype characteristics of common genotypes of LQTS can improve diagnostic accuracy, guide targeted genotyping, facilitate gene-specific treatment and improve risk stratification.

1. Effects of mutation type and location

The type and location of mutations may play an important role in the magnitude of repolarization abnormality, leading to occurrences of life threatening cardiac arrhythmias.

LQT1%The multi-center/multi-national LQTS registry study has shown that LQT1 patients with transmembrane mutations or mutations causing dominant-negative suppression of ion currents have longer QT intervals and a higher likelihood of developing life-threatening ventricular arrhythmias than patients with C-terminal mutations or mutations resulting in haploineffi-ciency. (100) It is possible that I_{Ks} channels with transmembrane mutations may have reduced responsiveness to regulator-adrenergic signaling of the ion conduction pathway with more impairment of shortening of the QTc with exercise related tachycardia than mutations in the C-terminus region. (101)

LQT2% Recently Shimizu et al (102) reported that LQT2 patients with missense mutations either at the transmembrane pore (S5-loop-S6) or at the N-terminus region of the hERG gene carry a higher risk of syncope or cardiac arrest (hazard ratio: 2.87 and 1.86, respectively). Those with the same type of mutation but at the transmembrane nonpore location have a lower risk. Among C-terminal mutations, LQT2 patients with non-missense mutations were at significantly higher risk than patients with missense mutations at the same region. Mutations located in a-helical domains are associated with a higher risk of cardiac events than mutations located in β -sheet domains.

LQT3% Previous studies suggest that the Δ KPQ mutation, located in the intracellular loops, and operating through both faster recovery from inactivation and an increase in residual sodium current, is associated with a significantly higher risk for cardiac events than the C-terminus D1790G mutation that has distinct biophysical function effects on steady-state inactivation and intracellular calcium homeostasis. (103) Depending on the outcome of the altered biophysical behavior of the sodium current, some SCN5A missense mutations can produce malignant phenotype and the severity is age-specific. (104)

- 2. Gene-specific triggers to cardiac events
- It has been well-documented that in LQT1 patients cardiac events are often triggered by exercise and swimming in particular. Of the patients who experienced cardiac events while swimming, 99% were LQT1. (105) In LQT2 patients only 13% occurred during exercise, most triggered by emotional stress.

Startle response with loud noise is hazardous to LQT2 patients. (106) In LQT3, sudden death most commonly occurs during sleep.

3. Genotype-specific ST-T-U wave patterns

The T wave morphology is abnormal in the majority of LQTS patients. Genotype-specific repolarization wave patterns have been identified in LQT1 to LQT3 and LQT7.

In 1995 Moss, et al (107) first identified three distinct T wave patterns associated with LQT1-3. Zhang et al (108) subsequently identified 10 ST-T wave repolarization patterns including 4 in LQT1, 4 in LQT2 and 2 in LQT3. LQT1 infants present a unique ST-T pattern classified as an infantile ST-T wave. The typical LQT1 ECG patterns in adults are featured by monomorphic T wave (Fig. 4) patterns in broad-based, normal-appearing or lateonset normal-appearing T waves. The normal appearing T wave is most common. Bifid T wave is a hallmark of LQT2 (Fig. 4). Bifid T waves can be obvious or subtle. Sometimes the second component of T wave is merged with U wave in precordial leads V2-3. Sometimes bifid T waves become very subtle that the top of the T wave become flat or rounded. In LQT3 the late-onset peaked/biphasic T waves are most common (Fig. 4). Typical ECG patterns are present in the majority of LQT1-3 mutation carriers. The ECG patterns are atypical in patients with compound mutations.

Characteristic ECG patterns are also found in ATS1 (LQT7). (80) The majority of *KCNJ2* mutation carriers present with abnormal T-U morphology featured by a prolonged terminal T wave downslope, wide T-U junction, biphasic and enlarged U waves. Frequent premature ventricular contractions (PVCs) in bigeminy and bidirectional ventricular tachycardia (VT) are common in ATS1. Interestingly, ventricular arrhythmias in ATS1 are originated from the left ventricle exclusively.

LQTS-specific arrhythmia

The arrhythmias in most of LQTS come in an "all-ornone" fashion. The signature arrhythmia of LQTS is TdP (Fig. 5). (5) Increased dispersion of ventricular repolarization and/or the development of early- or delayed-afterdepolarizations (EAD, DAD) by various causes can trigger TdP in the setting of an underlying long QT interval. (109, 110) The common ECG

Fig 4. Gene-specific T wave patterns in common LQTS genotypes. There are 10 ECG ST-T patterns typical to LQT1-3. Among them three representative ECG patterns are illustrated.

LQT1- The ECG tracing (25 mm/s) was taken from a 21year-old LQT1 female who had recurrent syncope and a family history of premature sudden death. The QT interval is markedly prolonged (QTc 540 ms) and the T wave is monomorphic and smooth (normal-appearing T wave). LQT2- The bifid T wave is of low amplitude and following a visible U wave. The QTc is markedly prolonged (521 ms). This tracing was taken from an asymptomatic 12-year-old girl carrying a KCNH2 mutation. LQT3- The Late-onset T waves visible amplitude with alternations were recorded from a symptomatic 6-year-old boy carrying a de novo SCN6A mutation. The QT is dramatically prolonged (QTc 590 ms).





Fig 5. The signature arrhythmia of LQTS The underlying cause of syncope in LQTS is usually torsade de pointes (TdP), a

torsade de pointes (TdP), a polymorphic ventricular tachycardia characterized by a pattern of twisting points. TdP in drug-induced LQTS usually starts with a short-long-short pattern of RR cycles. In this case, a short coupled PVC couplet is followed by a compensatory pause and then another PVC falls on the downslope of the T wave of the preceding sinus beat to start TdP. The QRS is wide with morphology and amplitude varied from beat to beat. The ORS axis switches in opposite direction around a virtual isoelectric baseline every 4-8 beats. In most cases TdP is short lived and terminates spontaneously. However it has a tendency to recur and sometimes it can degenerate into ventricular fibrillation, causing cardiac arrest and sudden death.

features of TdP include a markedly prolonged QT interval in the last sinus beat preceding the onset of the arrhythmia (pause-dependent), progressive twisting of the QRS complex polarity around an imaginary baseline, a complete 180-degree twist of the QRS complexes in 10 to 12 beats (150-300 bpm), and a changing amplitude of the QRS complexes in each cycle in a sinusoidal fashion.(10) Noda et al (111) observed three different initiating patterns in 24 congenital LQTS who had 111 TdPs recorded on ECG: (1) "pause-dependent" pattern (23 patients, 72 TdP, 65%); (2) "increased sinus rate" pattern (8 patients, 28 TdP, 25%) defined as a gradual increase in sinus rate with or without T-wave alternans; and (3) "changed depolarization" pattern (5 patients, 11 TdP, 10%) defined as a sudden long-coupled PVC or fusion beat followed by short-coupled PVC, suggesting differential mechanisms of initiation of TdP for each mode. Tan. et al (112) reported that pause-dependent TdP is predominantly seen in LQT2, which is also the case in drug-induced LQTS. In LQT1, a sudden intense adrenergic stimulation can precipitate TdP. In most cases TdP is short lived, terminating spontaneously thus may go unrecognized. However, it has a tendency to recur in rapid succession and can cause syncope. When TdP degenerates into VF, cardiac arrest or sudden death will be the outcome.

Natural history of the inherited LQTS

A much higher mortality reported in the earlier studies was drawn from smaller samples, mostly consisting of LQTS probands. A series of evidencebased studies conducted by International LQTS Registry investigators suggest that the risk of developing cardiac events in RWS is age-related, influenced by gender and the degree of QT prolongation. (113-115) The likelihood of developing cardiac events in infants is low in general. However, symptomatic LQTS babies bear a much higher risk of developing recurrent life-threatening events and the prognosis is poor, (116) especially in TS, JLNS and compound LQTS mutation carriers. In LQT1-3 children age 7 years and above, adolescents and young adults are the high risk groups. (117-118) Boys have higher risks than girls before age 13. Women at childbearing age, (114) especially those during postpartum, are more symptomatic. (119) A markedly prolonged QT interval is an independent risk factor for cardiac events. (120) In contrast to younger patients, a reduced frequency of cardiac events is seen in LQT1-3 patients age > 40. However, the mortality is still significantly higher than the age-matched non-LQTS elders.(121) Probands (the index case of each family) usually present with a much severe phenotype than the affected family members. (121) About 40-50% of LQT1-3 patients never develop cardiac events in their life time. Most asymptomatic LQTS individuals are not probands. Being asymptomatic does not guarantee his/her affected offspring will be risk free.(122) In RWS families, an offspring has a 50% probability of inheriting a mutant gene from his or her parent who carries a single mutation. In JLNS

families, congenital deafness occurs only in individuals carrying compound mutations (either heterozygous or homozygous) in *KCNQ1* (JLN1) or *KCNE1* (JLN2). (25) The prognosis is poor in patients with compound mutations. Their offspring have a 100% chance of inheriting a heterozygous mutation and is expected to present with a RWS phenotype. Heterozygous mutation carriers in JLNS families do not show deafness and usually present with a mild QT prolongation or even without a LQTS ECG phenotype (silent mutation carrier). Overall the cumulative rate of developing cardiac arrest and sudden death from birth to age 40 years is 5-8% in LQT1-3. Despite a high arrhythmia burden, the mortality in LQT7 is also low (3-5%).

Natural protection by genetic modifiers

Theoretically, dysfunction of the defective ion channel protein is the outcome of gene mutation. Why do 50% of LQTS mutation carriers never experience cardiac events in life? The mechanism of self-protection to prevent the worst outcome in LQTS is still largely unknown. More than 30% of the LQT2 mutations result in premature termination codons (PTCs). Degradation of PTCcontaining mRNA transcripts by nonsense-mediated mRNA decay (NMD) pathway can reduce mutant mRNA level and therefore eliminate the abundance of truncated proteins. Gong, et al (33) first reported in a large LQT2 family with C-terminus nonsense mutation that underwent NMD. All gene carriers except the proband presented with a mild phenotype. The proband on the other hand had two events of cardiac arrest at age of 30 to 40 years. The 1st event occurred when she was on a potassium depleting diet and the second happened while on a QT prolonging drug. It appears that risk factors such as the use QT prolonging drugs, electrolyte imbalance and gene-specific triggers can overwhelm the natural self-protection to initiate catastrophic arrhythmias in LQTS.

II. Acquired LQTS

Far more common than the congenital form is acquired LQTS that can also lead to TdP, cardiac arrest, and sudden death. They can be divided into several categories.

Drug-induced LQTS

Drug-induced LQTS is characterized by a prolonged QT interval after using one or more QT prolonging drug(s) and associated with an increased risk of TdP. (34) Most drugs that block the Kv11.1 (hERG) channel can prolong the QT interval. (7, 123, 124) However, only those producing inhomogeneous delay of repolarization are highly "TdP-genic". Many individuals developing drug-induced LQTS have underlying risk factors. A detailed drug list can be found at www.QTdrugs.org. Females are more susceptible than males to drug-induced TdP. (125, 126)

The molecular mechanisms of QT prolonging drugs include direct block of hERG channels (127) and

inhibition of hERG protein trafficking to the cell surface. (128)

- 1. Type-1 mechanism: direct I_{K_r} blocking is the most common, seen in a large number of structurally diverse therapeutic compounds including many antiarrhythmics, antihistamines, antipsychotics and antibiotics. The susceptibility of hERG blocking by small organic molecules is attributed to several unique structural features of the hERG channel. (129) 1) The high-affinity drug-binding site of hERG is located in the central cavity of the channel pathway (pore), which holds hydrated K⁺ ions on their approach to the selectivity filter once the cytoplasmic activation gate is opened. Consequently, most blockers bind only after the channel has opened, allowing their access to the central cavity of hERG. 2) Another crucial component of high-affinity drug binding originates from two concentric rings of aromatic amino acid side chains stacked on top of each other in the central cavity (Fig 6). Sanguinetti and Mitcheson (130) suggested that the surprising diversity and high affinity of hERG blockers depend to a large extent on hydrophobic interactions between the drug molecule and Phe656 residues and a cation- π interaction between a positively charged tertiary amine present in most blocking molecules and the π -electrons of Tyr652. Both Phe656 and Tyr652 are located in the S6 transmembrane domain and their side chains project directly into the central cavity of hERG. (131)
- 2. Type-2 mechanism: the inhibition of hERG protein trafficking (132) is seen in a number of agents, including Hsp90 function inhibitors such as geldanamycin, radiciol, 17-AGG, the metalloids arsenic trioxide and potassium antimony tartrate, as well as small organic molecules such as pentamidine, probucol and fluoxetine. The latter blocks hERG and disrupt its trafficking at the same time.

The severity of proarrhythmia at a given QT interval varies from drug to drug and from patient to patient. Unfortunately, the extent of QT prolongation and risk of TdP with a given drug may not be linearly related to the dose or plasma level of the drug as the metabolic factors are different individually (e.g. gender, electrolyte level). Furthermore, there is no linear relationship between the degree of drug induced QT prolongation and the likelihood of the development of TdP. The unintentional use of QT prolonging drugs has been recognized as a risk factor to trigger fatal cardiac events in patients with inherited LQTS. (31, 133)

Electrolyte imbalance

 Hypokalemia (serum level <3.5 mmol/L), a depletion or insulin-induced shifts of K⁺ from the extracellular compartment into cells, is the most common electrolyte imbalance related LQTS. Digestive or kidney disorders, and certain diuretics



Fig 6. Structural model of drug-binding sites in the hERG channel

Two of four subunits that form the pore and inner cavity of hERG are shown. The inner helices and loops extending from the pore helices to the selectivity filter form the inner cavity and drug-binding sites of hERG. Several features may help explain the nonspecific drug-binding properties of hERG. 1) The inner cavity of hERG is long and large, that can easily trap drugs to cause channel-drug interactions. The aromatic residues (not found in other K, channels) are critical drug-binding sites. 2) Other drug interaction sites are polar residues located close to the selectivity filter. Reproduced with permission from Michael Sanguinetti, PhD, the contributor of this illustration.

are common causes. (134) Serum potassium [K⁺]o is a key biologic modulator of cardiac ion currents. (129) For example, hERG potassium channel requires $[K^+]$ o for its membrane stability.(135) Under low [K⁺] o conditions, hERG channels in the plasma membrane are internalized and degraded. For a single hERG channel, the equilibrium between K⁺ bound and unbound state is [K⁺]o dependent. When [K⁺]o is decreased, the likelihood of a channel residing in K⁺ unbound state increases. Thus the plasma membrane density of hERG is strictly controlled by [K⁺]o. (135) In addition, K⁺ competes with Na⁺ for binding to the hERG external pore mouth. (136) Enhanced inactivation or increased inhibition by Na^+ under low $[K^+]o$ also contributes to a decreased I_{Kr} . At cellular level, reduced I_{K_r} by low [K⁺]o prolongs pha- se-3 action potential and facilitates EADs. (137) 3) Both external and internal $K^{\scriptscriptstyle +}$ modulates $I_{_{K1}}$ biophysic behavior. Inside of the cell, 70% of positive ions are K⁺. Cohen showed that a reduction in pipette $[K^+]$ from 145 to 25 mM, decreased the rate of activation of I_{K1} at a given voltage by several-fold, and reversed the voltage dependence of recovery from deactivation, so that the deactivation rate decreased with depolarization, and caused a positive shift in the midpoint of the activation curve of I_{κ_1} that was several fold smaller than the associated shift of the reversal potential. (138) Reduced I_{κ_1} prolongs the terminal phase of cardiac action potential and facilitates DADs. (139) The ECG presentation of hypokalemia is featured by low T wave amplitude, T-U merge, enlarged U wave,

and QTU prolongation. (140) Patients with severe hypokalemia are vulnerable to developing polymorphic PVCs and TdP/VF. (141) Hypoka-lemia is also an important risk factor for fatal arrhythmias in congential LQTS (33, 142) and individuals carrying functional cardiac ion channel gene SNPs.

2. Hypomagnesaemia (serum level <0.7 mmol/L) increases efflux of intracellular K⁺. The cell loses K⁺ which then is excreted by the kidneys, resulting in hypokalemia. Lack of magnesium inhibits the release of parathyroid hormone, which can result in hypoparathyroidism and hypocalcemia. Magnesium is an often overlooked electrolyte essential to all cells of all known living organisms. Magnesium plays an important role in enzymatic reactions and is critically involved in energy metabolism, glucose utilization, protein synthesis, fatty acid synthesis and breakdown, ATPase functions, and virtually all hormonal reactions. (143) Magnesium is also closely involved in maintaining cellular ionic balance through its association with sodium, potassium, and calcium, especially when transporting the ion across biological membranes. (144) i.e., magnesium is needed for the adequate function of the Na+/K+-ATPase pump to maintain the stability of resting potential. Lack of Mg²⁺ causes cells to depolarize easily and induce spontaneous arrhythmias. The clinical presentation of hypomagnesaemia is similar to that of hypo-kalemia, featuring QTU prolongation, TdP or even sudden death.(145, 146) About 10-20% of all hospital patients and 60-65% of patients in the intensive care unit have hypomagnesemia. Hypomagnesemia is under diagnosed since serum magnesium levels are not routinely tested.

Myocardial stunning

- 1. Neurogenic stunned myocardium is a reversible syndrome defined as myocardial injury and dysfunction occurring after diverse types of acute brain injury, as a result of imbalance of the autonomic nervous system. (147) The spectrum of observed cardiac abnormalities includes electrocardiographic changes, arrhythmia, myocardial necrosis, release of B-type natriuretic peptide, and both systolic and diastolic dysfunction of the left ventricle. Cardiac abnormalities have been associated with various CNS diseases, including trauma, ischemic stroke, and intracerebral hemorrhage. Neurogenic stunned myocardium is particularly common in patients with subarachnoid he-morrhage (SAH). (148) Less common etiologies include tumors, (149) electroconvulsive therapy, seizure disorders, and CNS infections such as meningitis. ECG abnormalities are more common in patients with intracranial hemorrhage (60%-70%) or SAH (40%-70%) than those with ischemic stroke (15%-40%). The most common ECG abnormality is QT prolongation, found in 45% to 71% of patients with SAH, 64% of patients with intraparenchymal hemorrhage, and 38% of patients with ischemic stroke. (147)
- 2. Tako-tsubo syndrome is another form of myocardial stunning precipitated by emotional or other types of stress but without acute brain injury. (148, 150) The clinical course is similar to neurogenic stunned myocardium, featuring reversible left ventricular apical ballooning associated with dynamic QT prolongation and diffuse T wave inversion. Patients with this syndrome are mostly elder females. Clinical presentation includes chest pain, pulmonary edema, and cardiogenic shock. There is a mildly elevated serum troponin I level, (151) but angiographic evidence of clinically significant coronary disease is uncommon. Left ventricular dysfunction is usually resolved within a few weeks to a few months.

Unlike congenital and drug-induced LQTS, the dynamic changes of marked QT prolongation and diffuse T wave inversion seen in myocardial stunning may be largely due to an altered recovery sequence corresponding to the marked regional wall motion abnormalities. Such alterations may not be always associated with increased transmural dispersion of repolarization (TDR). This hypothesis may explain the fact that TdP is infrequent though it can occur and cause sudden death. Once the primary cause is removed most patients experience a benign clinical course.

Oxidation of hERG channels

Under pathologic conditions (i.e. reperfusion after acute ischemia) hERG channels are exposed to a burst of excess of reactive species including reactive oxygen and nitrogen. hERG channels can be altered by reactive species resulting in redox-dependent inactivation of α - and β -subunits channel complexes. Dysfunction of hERG channels caused by methionine oxidation is likely to occur during oxidative stress mediated by an imbalance of oxidation and elimination. hERG oxidation may be an important mechanism of acquired LQTS under oxidative stress. (152)

Other causes

A prolonged QT interval can be secondary to cardiomyopathies (hypertrophic, dilated or arrhythmogenic, etc), mitral valve prolapse, congestive heart failure, hypertension, coronary artery disease, complete heart block, (153, 154) Kawasaki syndrome, myocarditis, diabetes, anorexia nervosa, and hepatic impairment. Cardiac ion channel remodeling due to the primary condition may have resulted in a delayed repolarization. Many patients in this group also take QT prolonging drugs, with a further increased risk of sudden death.

Female gender is a risk factor for various acquired LQTS induced TdP. Female predominance in bradycardia- and electrolyte imbalance-induced LQTS has also been reported (125, 155, 156)

Genetic predisposition to acquired LQTS

Like *KCNE1* (157) and *KCNE2*, *KCNE3* is the third modifier gene in *KCNE* family expressed in cardiac myocytes and interacts with *KCNQ1* to change channel properties. Two *KCNE3* missense mutations located in the N- and C-terminal domains are linked to drug induced- and hypokalemia-induced TdP. (69, 158) Silent LQTS gene carriers (142, 159) and certain genetic modifiers such as some of the functional SNPs, (160-162) can affect the biophysical properties of cardiac ion channels, (163) resulting in a hidden condition called "reduced repolarization reserve" (164) which can be surfaced to the extent of disrupting cardiac ion channel function in the presence of QT prolonging drugs, electrolyte imbalance, ischemia or many other acquired conditions.(69, 157, 163, 165-168)

Cellular mechanism and experimental models of LQTS Prolongation of QT interval, the time course between ventricular depolarization and repolarization, on the surface ECG is caused by lengthening the APD of ventricular myocytes. Prolongation of the QT interval can occur as a consequence of congenital defects or in response to drugs that prolong the APD via a reduction in I_{Ks} , I_{Kr} , or I_{K1} ; or an increase in I_{Ca} or late I_{Na} . Though the inherited forms of the LQTS are phenotypically and genotypically diverse, they all have one thing in

common: a prolonged QT interval associated with an increased risk of arrhythmic sudden death. (21)

Work by Antzelevitch and his associates (169-171) have demonstrated that the ventricular myocardium is composed of at least three electrophysiologically and functionally distinct cell types: epicardial, mid-myocardial (M), and endocardial cells. The hallmark of the M cell is the ability of its action potential to prolong more than that of epicardium or endocardium in response to a slowing of rate. This feature of the M cell is due to weaker repolarizing current during phases 2 and 3 secondary to a smaller $I_{\rm Ks}$ and a larger late $I_{\rm Na}$ and $I_{\rm Na-Ca}$ compared with epicardial and endocardial cells. (172)

Under normal and most long QT conditions, the epicardial cell is the earliest to repolarize and the M cell is often the last. In an arterially perfused left ventricular wedge preparation with bipolar ECG leads placed across the transmural ventricular wall, repolarization of the epicardial action potential is coincident with the peak of the T wave and repolarization of the M cells coincides with the end of the T wave. (19) Thus, the repolarization of the M cells usually determines the QT interval. The interval between the peak and end of the T wave (Tpeak-Tend) has been suggested to provide an index of transmural dispersion of repolarization. (19) Amplification of spatial dispersion of repolarization within the ventricular myocardium has been identified as the principal arrhythmogenic substrate in both acquired and congenital LQTS. Accentuation of spatial dispersion, typically secondary to an increase of transmural, trans-septal or apico-basal dispersion of repolarization, and the development of EADs underlie the substrate and trigger for the development of TdP observed under LQTS conditions. Models of LQT1-3 and 7 have been developed using the canine arterially perfused left ventricular wedge preparations. (19, 139, 173-175) In LQT1-3 models preferential prolongation of the M cell APD can lead to an increase in the QT interval as well as an increase in transmural dispersion of repolarization, which contributes to the development of TdP. (21)Morita et al (139) demonstrated that in the LQT7 model under low [K⁺]o conditions the delayed late phase-3 repolarization and DADs generate U waves. Additional I_{κ_1} blockage by cesium chloride plus sympathetic activation by isoproterenol increased frequency of DADs. Alternating DADs at two foci resulted in bidirectional VTs.

Diagnosis of LQTS

Since treatment strategies are different, acquired and congenital forms should be differentiated. In an acquired form, the primary disease and/or the cause of QT prolongation can be identified. Drugs, hypokalemia and hypomagnesaemia-induced LQTS are often associated with dynamic QTU prolongation and T-U merge with a high tendency to the development of TdP. Acquired LQTS usually do not have the characteristic ECG patterns seen in the congenital forms. The clinical diagnosis of congenital LQTS on the other hand primarily depends on the ECG findings, family history of unexplained sudden death, blood relative(s) diagnosed with LQTS, and history of cardiac events. Electrophysiological studies are not helpful in diagnosing LQTS since TdPs are mostly non-inducible. Commercial genetic testing is fast, accurate and reliable in the United States. Considering the cost, it should be used based on appropriate indications. In 1993 Schwartz et al (2) suggested incorporating both clinical and ECG findings in probability-based diagnostic criteria for diagnosing LQTS. The maximum score is 9, and a score of 3 or more indicates a high probability of LQTS. The criteria are as follows:

ECG findings (without medications or disorders known to affect ECG features)

QT corrected for heart rate (QTc), calculated using Bazett's formula, of more than 480 milliseconds (ms) – 3 points

QTc of 460-470 ms – 2 points

- QTc of 450 ms in male patients 1 point
- Torsade de pointes (mutually exclusive) 2 points T-wave alternans – 1 point
- Notched T wave in 3 leads 1 point

Low heart rate for age (ie, resting heart rate below the second percentile for age) -0.5 points

Clinical history

Syncope with stress (mutually exclusive) -2 points Syncope without stress -1 point

Congenital deafness – 0.5 points

Family history (The same family member cannot be counted in both categories)

Family member with definite long QT syndrome – 1 point

Unexplained sudden cardiac death (age <30 y) in an immediate family member – 0.5 points

These criteria provide a quantitative approach to the diagnosis of LQTS by allocating numerical points to clinical features, family history, and ECG findings and divide the possibility of LQTS into low, intermediate, and high probability ranges. Based on experience and available information from genotype-phenotype correlation studies, we have focused on the following aspects which can help increase the diagnostic accuracy.

Syncope

The most common cardiac event in LQTS is syncope. LQTS-related syncope is often confused with the common faint, known as vasovagal or neurocardiogenic syncope. A careful evaluation of factors surrounding syncope can help the differentiation.(176) The LQTS syncope is usually precipitous without warning. It occurs during or just after physical exertion, emotional excitement or sudden auditory arousal (such as a doorbell or alarm clock), but may occur during sleep or at rest. Conversely, in vasovagal syncope, usually there are warning symptoms, such as dizziness, blurring or blackening vision, tingling or sweating for seconds or even minutes prior to syncope. Also, a precipitating event is usually present, commonly pain, injury, nausea, or unpleasant or stressful experience.

Recurrent syncope is also confused by epilepsy, due to a sudden surge of electrical activity in the brain that results in complete loss of consciousness. Many electroencephalography (EEG) laboratories routinely monitor the ECG during EEG recordings. This has proven useful in detecting cardiac rhythm disorders which may or may not account for the patient's symptoms but should result in referral to a cardiologist. Recent studies suggest that LQTS patients, especially LQT2, can develop epilepsy because on channel proteins are not only expressed in the heart but also in the brain. (177)

Family investigation

Unexplained syncope or premature sudden death especially in a child or young adult should raise suspicion of a channelopathy. ECG screening of family members to determine whether a QT prolongation can be found in blood relatives is very important. Since most cases of congenital LQTS are inherited, and since up to 50% of sudden death occurred as the first symptom, once a diagnosis is established in a patient, conducting a family search starting from the 1st degree blood relatives is highly recommended.(97)

ECG evaluation

- 1. QT interval making a correct QT measurement is extremely important in the LQTS diagnosis. (178, 179) Sometimes the QT readings on automatic computation can be misleading especially in individuals with a complicated T wave morphology or in the presence of a prominent U wave or TU merge. (180) In general, U wave should be excluded in QT measurement. (178) Manually measuring the QT interval to validate the machine reading is necessary for LQTS diagnosis. Among many QT formulae proposed Bazett's (QTc=QT/ \sqrt{RR}) is the most used among clinicians. When the heart rate is in the physiologic range, Bazett's formula is reliable and the easiest in clinical settings and for drug efficacy evaluation and follow-up studies. To avoid false positives averaging R-R interval of 10 seconds or a minimum 2-3 consecutive beats is recommended for QTc calculations. Although the normal upper limit is 440 ms, there is a considerable overlap of QTc in the range of 450-470 ms between normal and LQTS subjects, complicating the ECG diagnosis.(4) Women have longer QT intervals than men. (181) The cut points of $QTc \ge 470$ ms in males and ≥ 480 ms in females are insensitive but highly specific in LQTS diagnosis.(4)Additional evaluation is required in subjects with a borderline QT interval.
- 2. T wave morphology-diagnosing LQTS should not be based upon the QT reading alone. A prolonged QT interval in LQTS is often associated with

abnormal T wave morphology. (107) Recognizing the gene-specific ST-T-U morphology in common genotypes can increase diagnostic accuracy. (97, 108, 182) Sometimes it helps to predict the underlying genotype. (80, 108, 183)

3. Serial ECGs - it is helpful to those showing a bordering QT prolongation at the initial screening since QT interval and T wave morphology may vary from time to time.(97) Serial ECGs can increase the chance of capturing a typical T wave pattern and/or a diagnostic QT interval. (97, 184)

Stress testing

- 1. Exercise testing For subjects with a borderline QT interval and/or atypical T wave patterns exercise testing can be of help.(30, 185, 186) In LQT1 QTc lengthening and broad-based T wave occur in the early recovery stages. In contrast, QT shortens and the T wave normalizes in the peak exercise and early recovery stages in LQT3. In LQT2 the T wave often becomes more bifid in the later stages of recovery. The QT maximum that is diagnostic can be obtained from exercise testing and the QT response to exercise can distinguish LQT1, LQT2 and LQT3 (187) in most cases. For many years the "Vincent Bicycle Protocol" and/or a treadmill with a modified Bruce protocol have been used in testing adults and children. It is a safe, reliable and easy to reproduce in LQTS diagnosis and follow-up (unpublished information by the author). Exercise testing is also helpful in beta-blocker efficacy evaluations.
- 2. Drug-challenge tests Epinephrine QT stress testing is also an effective diagnostic tool to unmask concealed LQTS, particularly LQT1. Unique responses have also been observed in patients with LQT2 and LQT3, making this test useful in the diagnostic work-up of LQTS. (188, 189)

Genetic testing and functional expression

Though LQTS is mostly a single gene channelopathy, most mutations are "private" or "family-specific" and few are hot-spot mutations. (96, 190) To date, over 1,200 mutations in at least 12 genes have been identified worldwide. It is impractical, time consuming and costly to screen all known genes by a research laboratory. In a large cohort of unrelated patients referred for LQTS genetic testing, Tester, et al (99) reported that the clinical phenotype strongly correlated with the likelihood of elucidating a pathogenic mutation with the cardiac channel gene screen. Since the vast majority of LQTS patients with mutations identified are LQT1-3, and gene-specific ECG patterns are present in the majority of LQT1-3 patients that can be easily recognized by clinicians, an ECG-based genotype prediction can provide a guide for targeted gene search that has proven cost-effective in developing countries. (183) In the United States, LQTS-gene screening is available in commercial genetic laboratories but frequently not paid by

commercial insurers. Deletion and duplication should be screened in patients with negative findings of known gene sequencing. (98) Using advanced systems, commercial labs can provide genotyping results of known genes within 1-2 months. The chance of identifying a mutation is 75% among those with a clear LQTS phenotype. Overall, genetic testing is an important aspect in LQTS diagnosis. (23) In the future, genotypic determination of LQTS patients and their family members will hopefully lead to improved gene-specific prognostic determinations and therapeutic interventions.

Treatment of LQTS

The ultimate goal of LQTS treatment is to prevent cardiac arrest and sudden death. (191) In acquired forms, removing the causative agent is the key. Magnesium can suppress TdP effectively. (192) In congenital LQTS, avoiding the use of QT prolonging drugs, keeping electrolytes in balance and avoiding gene-specific situational or environmental triggers can lower the risk of cardiac events. Depending on the genotype and severity of phenotype, treatment strategy may vary. In general, medications, cardiac devices, and surgical approaches are the therapeutic choices for congenital LQTS.

β-blocker therapy

Among all the drugs β -blocker therapy is the only one that has provided significant morbidity and mortality benefits in patients who have sustained acute myocardial infarction. β -blocker therapy is most effective in preventing cardiac events in LQTS and has served as the mainstay choice for the last three decades. Yearly follow-up to justify the dose or the body weight increase is recommended for children especially teenagers. Compliance is the key to effective therapy. In a recent retrospective study by Vincent, et al (31) β blockers were highly effective in reduced cardiac events in LQT1 patients. They suggested that β -blockers should be administered at diagnosis and ideally before the preteen years. Propranolol (48%) and nadolol (36%)were most frequently prescribed β -blockers. The mean daily dose was 2.2±1.1 mg/kg for propranolol, and 1.7 ± 0.79 mg/kg for nadolol. In this cohort noncompliance and use of QT-prolonging drug were responsible for almost all life-threatening cardiac events. In general, the protection of β -blockers is weighted by genotype as LQT1>LQT2>LQT3. A recent computational study indicated that high dose β -blocker is likely protective in LQT3 as well. However, patients with the following conditions are not suitable for β -blocker therapy : 1) marked bradycardia especially associated with sinus node dysfunction; 2) intolerance to β -blockers; and 3) symptomatic despite β -blocker medications. Certain genetic modifiers may affect the effectiveness of β -blocker therapy. (193-197)

Potassium supplements

Potassium supplements may be helpful in people with certain forms of long QT syndrome, such as LQT2 (198-199) and Andersen-Tawil syndrome.(79) Magnesium supplement is suitable to patients with hypomagnesaemia-induced LQTS.

Implantable Cardioverter Defibrillators (ICD)

ICDs automatically sense life-threatening rhythms and deliver electrical therapy or life-saving shock directly to the heart. This therapy has clearly been shown to be the most effective in aborting arrhythmic sudden death. (191) How much the benefit outweighs potential risks in terms of morbidity and quality of life is less clear. An ICD or ICD/pacemaker combination may be used in the following cases: 1) Medications do not work or the individual has unacceptable side effects from the medications. 2) The patient's first symptom is an aborted sudden death. Pacemaker or ICD/pacer is suitable to symptomatic patients with severe bradycardia or sinus node dysfunction.

Left cardiac sympathetic denervation (LCSD)

In 1971 Moss used LCSD to treat a LQTS patient who was refractory to pharmacologic therapy. (200) The methodology has improved over time. The high thoracic left sympathectomy has been adopted, in which the lower part of the left stellate ganglion together with the first four or five left thoracic ganglia all removed. This procedure produces an adequate cardiac sympathetic denervation and is associated with a very low incidence of Horner's syndrome. LCSD is known to raise the threshold for ventricular fibrillation and reduce the arrhythmias associated with acute myocardial ischemia in animal models without reducing heart rate or impairing myocardial contractility. A significant protective effect of LCSD among high-risk patients with LQTS was shown by Schwartz et al. (201, 202) Li (203) took a less invasive approach by videoscopic denervation, which has shown the promising result. Videoscopic denervation can provide faster recovery with much less surgical trauma than the traditional LCSD. (204, 205)

Prognosis

Once diagnosed LQTS can be effectively treated and sudden death can be prevented. Not every LQTS needs an ICD especially since up to 50% of gene mutation carriers never experience cardiac events. Risk stratification of the individual patient optimizes appropriate and individualized therapy and clinical outcomes.

Conflict of Interest

None of the authors have financial or other conflict of interest to disclose

REFERENCES

1. Jervell A, Lange-Nielsen F. Congenital deaf-mutism, functional heart disease with prolongation of the Q-T interval and sudden death. Am Heart J 1957;54:59-68.

2. Schwartz PJ, Moss AJ, Vincent GM, Crampton RS. Diagnostic criteria for the long QT syndrome. An update. Circulation 1993;88:782-4.

Moss AJ. Prolonged QT-interval syndromes. JAMA 1986;256:2985-7.
 Vincent GM, Timothy KW, Leppert M, Keating M. The spectrum of symptoms and QT intervals in carriers of the gene for the long-QT syndrome. N Engl J Med 1992;327:846-52.

5. Jackman WM, Clark M, Friday KJ, Aliot EM, Anderson J, Lazzara R. Ventricular tachyarrhythmias in the long QT syndromes. Med Clin North Am 1984;68:1079-109.

6. Jackman WM, Friday KJ, Anderson JL, Aliot EM, Clark M, Lazzara R. The long QT syndromes: a critical review, new clinical observations and a unifying hypothesis. Prog Cardiovasc Dis 1988;31:115-72.

7. Roden DM. Cellular basis of drug-induced torsades de pointes. Br J Pharmacol 2008;154:1502-7.

8. Viskin S, Alla SR, Barron HV, Heller K, Saxon L, Kitzis I, et al. Mode of onset of torsade de pointes in congenital long QT syndrome. J Am Coll Cardiol 1996;28:1262-8.

9. Napolitano C, Priori SG, Schwartz PJ. Torsade de pointes. Mechanisms and management. Drugs 1994;47:51-65.

10. Drew BJ, Ackerman MJ, Funk M, Gibler WB, Kligfield P, Menon V, et al; American Heart Association Acute Cardiac Care Committee of the Council on Clinical Cardiology, the Council on Cardiovascular Nursing, and the American College of Cardiology Foundation. Prevention of Torsade de Pointes in Hospital Settings. A Scientific Statement From the American Heart Association and the American College of Cardiology Foundation. Circulation 2010;121:1047-60.

11. Viskin S. Long QT syndromes and torsade de pointes. Lancet 1999;354:1625-33.

12. Schwartz PJ, Priori SG, Spazzolini C, Moss AJ, Vincent GM, Napolitano C, et al. Genotype-phenotype correlation in the long-QT syndrome: gene-specific triggers for life-threatening arrhythmias. Circulation 2001;103:89-95.

13. Ackerman MJ, Tester DJ, Porter CJ. Swimming, a gene-specific arrhythmogenic trigger for inherited long QT syndrome. Mayo Clin Proc 1999;74:1088-94.

14.Wilde AA, Jongbloed RJ, Doevendans PA, Duren DR, Hauer RN, van Langen IM, et al. Auditory stimuli as a trigger for arrhythmic events differentiate HERG-related (LQTS2) patients from KVLQT1-related patients (LQTS1). J Am Coll Cardiol 1999;33:327-32.

15. Ackerman MJ. The long QT syndrome: ion channel diseases of the heart. Mayo Clin Proc 1998;73:250-69.

16. Antzelevitch C. Molecular biology and cellular mechanisms of Brugada and long QT syndromes in infants and young children. J Electrocardiol 2001;34:177-81.

17. Wilde AA, Veldkamp MW. Ion channels, the QT interval, and arrhythmias. Pacing Clin Electrophysiol 1997;20:2048-51.

18.Spooner PM, Albert C, Benjamin EJ, Boineau R, Elston RC, George AL Jr, et al. Sudden cardiac death, genes, and arrhythmogenesis : consideration of new population and mechanistic approaches from a national heart, lung, and blood institute workshop, part I. Circulation 2001;103:2361-4.

19. Yan GX, Antzelevitch C. Cellular basis for the normal T wave and the electrocardiographic manifestations of the long-QT syndrome. Circulation 1998;98:1928-36.

20. Antzelevitch C. Cellular basis and mechanism underlying normal and abnormal myocardial repolarization and arrhythmogenesis. Ann Med 2004;36:5-14.

21. Antzelevitch C. Ionic, molecular, and cellular bases of QT-interval prolongation and torsade de pointes. Europace 2007;9:iv4-15.

22. Schwartz PJ, Stramba-Badiale M, Crotti L, Pedrazzini M, Besana A, Bosi G, et al. Prevalence of the congenital long-QT syndrome. Circulation 2009;120:1761-7.

23. Kapa S, Tester DJ, Salisbury BA, Harris-Kerr C, Pungliya MS, Alders M, et al. Genetic testing for long-QT syndrome: distinguishing pathogenic mutations from benign variants. Circulation 2009;120:1752-60.

24. Nerbonne JM, Kass RS. Molecular physiology of cardiac repolarization. Physiol Rev 2005;85:1205-53.

25. Schwartz PJ, Spazzolini C, Crotti L, Bathen J, Amlie JP, Timothy K, et al. The Jervell and Lange-Nielsen syndrome: natural history, molecular basis, and clinical outcome. Circulation 2006;113:783-90.
26. Wang Q, Curran ME, Splawski I, Burn TC, Millholland JM, VanRaay TJ, et al. Positional cloning of a novel potassium channel gene: KVLQT1 mutations cause cardiac arrhythmias. Nat Genet 1996;12:17-23.

27. Sanguinetti MC, Curran ME, Zou A, Shen J, Spector PS, Atkinson DL, et al. Coassembly of K(V)LQT1 and minK (IsK) proteins to form cardiac I(Ks) potassium channel. Nature 1996;384:80-3.

28. Splawski I, Shen J, Timothy KW, Lehmann MH, Priori S, Robinson JL, et al. Spectrum of mutations in long-QT syndrome genes. KVLQT1, HERG, SCN5A, KCNE1, and KCNE2. Circulation 2000;102:1178-85.

29. Wang Z, Tristani-Firouzi M, Xu Q, Lin M, Keating MT, Sanguinetti MC. Functional effects of mutations in KvLQT1 that cause long QT syndrome. J Cardiovasc Electrophysiol 1999;10:817-26.

30. Vincent GM, Jaiswal D, Timothy KW. Effects of exercise on heart rate, QT, QTc and QT/QS2 in the Romano-Ward inherited long QT syndrome. Am J Cardiol 1991;68:498-503.

31. Vincent GM, Schwartz PJ, Denjoy I, Swan H, Bithell C, Spazzolini C, et al. High efficacy of beta-blockers in long-QT syndrome type 1: contribution of noncompliance and QT-prolonging drugs to the occurrence of beta-blocker treatment "failures". Circulation 2009;119:215-21.

32. Curran ME, Splawski I, Timothy KW, Vincent GM, Green ED, Keating MT. A molecular basis for cardiac arrhythmia: HERG mutations cause long QT syndrome. Cell 1995;80:795-803.

33. Gong Q, Zhang L, Vincent GM, Horne BD, Zhou Z. Nonsense mutations in hERG cause a decrease in mutant mRNA transcripts by nonsense-mediated mRNA decay in human long-QT syndrome. Circulation 2007;116:17-24.

34. Zhou Z, Gong Q, Epstein ML, January CT. HERG channel dysfunction in human long QT syndrome. Intracellular transport and functional defects. J Biol Chem1998;273:21061-6.

35. Zhou Z, Gong Q, January CT. Correction of defective protein trafficking of a mutant HERG potassium channel in human long QT syndrome. Pharmacological and temperature effects. J Biol Chem1999;274:31123-6.

36. January CT, Gong Q, Zhou Z. Long QT syndrome: cellular basis and arrhythmia mechanism in LQT2. J Cardiovasc Electrophysiol 2000;11:1413-8.

37. Anderson CL, Delisle BP, Anson BD, Kilby JA, Will ML, Tester DJ, et al. Most LQT2 mutations reduce Kv11.1 (hERG) current by a class 2 (trafficking-deficient) mechanism. Circulation 2006;113:365-73.

38. Wang Q, Shen J, Li Z, Timothy K, Vincent GM, Priori SG, et al. Cardiac sodium channel mutations in patients with long QT syndrome, an inherited cardiac arrhythmia. Hum Mol Genet 1995;4:1603-7.

39. Wang Q, Li Z, Shen J, Keating MT. Genomic organization of the human SCN5A gene encoding the cardiac sodium channel. Genomics 1996;34:9-16.

40. George AL Jr. Inherited disorders of voltage-gated sodium channels. J Clin Invest 2005;115:1990-9.

41. George AL Jr. Hereditary dysfunction of voltage-gated sodium channels: from clinical phenotype to molecular mechanisms. Nephrol Dial Transplant 1996;11:1730-7.

42. George AL Jr, Knittle TJ, Tamkun MM. Molecular cloning of an atypical voltage-gated sodium channel expressed in human heart and uterus: evidence for a distinct gene family. Proc Natl Acad Sci U S A 1992;89:4893-7.

43. Zimmer T, Surber R. SCN5A channelopathies-an update on mutations and mechanisms. Prog Biophys Mol Biol 2008;98:120-36.

44. Zimmer T, Bollensdorff C, Haufe V, Birch-Hirschfeld E, Benndorf K. Mouse heart Na+ channels: primary structure and function of two isoforms and alternatively spliced variants. Am J Physiol Heart Circ Physiol 2002;282:H1007-17.

45. Tester DJ, Ackerman MJ. Sudden infant death syndrome: how significant are the cardiac channelopathies? Cardiovasc Res 2005;67:388-96.

46. Weese-Mayer DE, Ackerman MJ, Marazita ML, Berry-Kravis EM. Sudden Infant Death Syndrome: review of implicated genetic factors. Am J Med Genet A 2007;143A:771-88.

47. Chen Q, Kirsch GE, Zhang D, Brugada R, Brugada J, Brugada P, et al. Genetic basis and molecular mechanism for idiopathic ventricular fibrillation. Nature 1998;392:293-296.

48. Wang DW, Makita N, Kitabatake A, Balser JR, George AL Jr. Enhanced Na(+) channel intermediate inactivation in Brugada syndrome. Circ Res 2000;87:E37-43.

49. Schott JJ, Alshinawi C, Kyndt F, Probst V, Hoorntje TM, Hulsbeek M, et al. Cardiac conduction defects associate with mutations in SCN5A. Nat Genet 1999;23:20-1.

50. Tan HL, Bink-Boelkens MT, Bezzina CR, Viswanathan PC, Beaufort-Krol GC, van Tintelen PJ, et al. A sodium-channel mutation causes isolated cardiac conduction disease. Nature 2001;409:1043-7.
51. Wang DW, Viswanathan PC, Balser JR, George AL, Jr., Benson DW. Clinical, genetic, and biophysical characterization of SCN5A mutations associated with atrioventricular conduction block. Circulation 2002;105:341-6.

52. Benson DW, Wang DW, Dyment M, Knilans TK, Fish FA, Strieper MJ, et al. Congenital sick sinus syndrome caused by recessive mutations in the cardiac sodium channel gene (SCN5A). J Clin Invest 2003;112:1019-28.

53. Takehara N, Makita N, Kawabe J, Sato N, Kawamura Y, Kitabatake A, et al. A cardiac sodium channel mutation identified in Brugada syndrome associated with atrial standstill. J Intern Med 2004;255:137-42.

54. Rossenbacker T, Carroll SJ, Liu H, Kuiperi C, de Ravel TJ, Devriendt K, et al. Novel pore mutation in SCN5A manifests as a spectrum of phenotypes ranging from atrial flutter, conduction disease, and Brugada syndrome to sudden cardiac death. Heart Rhythm 2004;1:610-5.

55. Makiyama T, Akao M, Shizuta S, Doi T, Nishiyama K, Oka Y, et al. A novel SCN5A gain-of-function mutation M1875T associated with familial atrial fibrillation. J Am Coll Cardiol 2008;52:1326-34.

56. Darbar D, Kannankeril PJ, Donahue BS, Kucera G, Stubblefield T, Haines JL, et al. Cardiac sodium channel (SCN5A) variants associated with atrial fibrillation. Circulation 2008;117:1927-35.

57. Olson TM, Michels VV, Ballew JD, Reyna SP, Karst ML, Herron KJ, et al. Sodium channel mutations and susceptibility to heart failure and atrial fibrillation. JAMA 2005;293:447-54.

58. Shi R, Zhang Y, Yang C, Huang C, Zhou X, Qiang H, et al. The cardiac sodium channel mutation delQKP 1507-1509 is associated with the expanding phenotypic spectrum of LQT3, conduction disorder, dilated cardiomyopathy, and high incidence of youth sudden death. Europace 2008;10:1329-35.

59. Eckardt L, Kirchhof P, Loh P, Schulze-Bahr E, Johna R, Wichter T, et al. Brugada syndrome and supraventricular tachyarrhythmias: a novel association? J Cardiovasc Electrophysiol 2001;12:680-5.

60. Remme CA, Wilde AA. SCN5A overlap syndromes: no end to disease complexity? Europace 2008;10:1253-5.

61. Schott JJ, Charpentier F, Peltier S, Foley P, Drouin E, Bouhour JB, et al. Mapping of a gene for long QT syndrome to chromosome 4q25-27. Am J Hum Genet 1995;57:1114-22.

62. Mohler PJ, Bennett V. Ankyrin-based cardiac arrhythmias: a new class of channelopathies due to loss of cellular targeting. Curr Opin Cardiol 2005;20:189-93.

63. Mohler PJ, Le Scouarnec S, Denjoy I, Lowe JS, Guicheney P, Caron L, et al. Defining the cellular phenotype of "ankyrin-B syndrome" variants: human ANK2 variants associated with clinical phenotypes display a spectrum of activities in cardiomyocytes. Circulation 2007;115:432-41.

64. Mohler PJ, Schott JJ, Gramolini AO, Dilly KW, Guatimosim S, duBell WH, Song LS, Haurogne K, Kyndt F, Ali ME, Rogers TB, Lederer WJ, Escande D, Le Marec H, Bennett V. Ankyrin-B mutation causes type 4 long-QT cardiac arrhythmia and sudden cardiac death. Nature 2003;421:634-9.

65. Mohler PJ, Splawski I, Napolitano C, Bottelli G, Sharpe L, Timothy K, et al. A cardiac arrhythmia syndrome caused by loss of ankyrin-B function. Proc Natl Acad Sci U S A. 2004;101:9137-42.

66. Splawski I, Tristani-Firouzi M, Lehmann MH, Sanguinetti MC, Keating MT. Mutations in the hminK gene cause long QT syndrome and suppress IKs function. Nat Genet 1997;17:338-40.

67. Ohno S, Zankov DP, Yoshida H, Tsuji K, Makiyama T, Itoh H, et al. N- and C-terminal KCNE1 mutations cause distinct phenotypes of long QT syndrome. Heart Rhythm 2007;4:332-40.

68. Harmer SC, Tinker A. The role of abnormal trafficking of KCNE1 in long QT syndrome 5. Biochem Soc Trans 2007;35:1074-6.

69. Nishio Y, Makiyama T, Itoh H, Sakaguchi T, Ohno S, Gong YZ, et al. D85N, a KCNE1 polymorphism, is a disease-causing gene variant in long QT syndrome. J Am Coll Cardiol 2009;54:812-9.

70. Abbott GW, Sesti F, Splawski I, Buck ME, Lehmann MH, Timothy KW, et al. MiRP1 forms IKr potassium channels with HERG and is associated with cardiac arrhythmia. Cell 1999;97:175-87.

71. Gordon E, Panaghie G, Deng L, Bee KJ, Roepke TK, Krogh-Madsen T, et al. A KCNE2 mutation in a patient with cardiac arrhythmia induced by auditory stimuli and serum electrolyte imbalance. Cardiovasc Res 2008;77:98-106.

72. Isbrandt D, Friederich P, Solth A, Haverkamp W, Ebneth A, Borggrefe M, et al. Identification and functional characterization of a novel KCNE2 (MiRP1) mutation that alters HERG channel kinetics. J Mol Med 2002;80:524-32.

73. Andersen ED, Krasilnikoff PA, Overvad H. Intermittent muscular weakness, extrasystoles, and multiple developmental anomalies. A new syndrome? Acta Paediatr Scand 1971;60:559-64.

74. Tawil R, Ptacek LJ, Pavlakis SG, DeVivo DC, Penn AS, Ozdemir C, et al. Andersen's syndrome: potassium-sensitive periodic paralysis, ventricular ectopy, and dysmorphic features. Ann Neurol 1994;35:326-30.

75. Andelfinger G, Tapper AR, Welch RC, Vanoye CG, George AL Jr, Benson DW. KCNJ2 mutation results in Andersen syndrome with sex-specific cardiac and skeletal muscle phenotypes. Am J Hum Genet 2002;71:663-8.

76. Dhamoon AS, Pandit SV, Sarmast F, Parisian KR, Guha P, Li Y, et al. Unique Kir2.x properties determine regional and species differences in the cardiac inward rectifier K+ current. Circ Res 2004;94:1332-9.

77. Lopatin AN, Nichols CG. Inward rectifiers in the heart: an update on I(K1). J Mol Cell Cardiol 2001;33:625-38.

78. Wang Z, Yue L, White M, Pelletier G, Nattel S. Differential distribution of inward rectifier potassium channel transcripts in human atrium versus ventricle. Circulation 1998;98:2422-8.

79. Tristani-Firouzi M, Jensen JL, Donaldson MR, Sansone V, Meola G, Hahn A, et al. Functional and clinical characterization of KCNJ2 mutations associated with LQT7 (Andersen syndrome). J Clin Invest 2002;110:381-8.

80. Zhang L, Benson DW, Tristani-Firouzi M, Ptacek LJ, Tawil R, Schwartz PJ, et al. Electrocardiographic features in Andersen-Tawil syndrome patients with KCNJ2 mutations: characteristic T-U-wave patterns predict the KCNJ2 genotype. Circulation 2005;111:2720-6.
81. Splawski I, Timothy KW, Sharpe LM, Decher N, Kumar P, Bloise R, et al. Ca(V)1.2 calcium channel dysfunction causes a multisystem disorder including arrhythmia and autism. Cell 2004;119:19-31.

82. Splawski I, Timothy KW, Decher N, Kumar P, Sachse FB, Beggs AH, et al. Severe arrhythmia disorder caused by cardiac L-type calcium channel mutations. Proc Natl Acad Sci U S A 2005;102:8089-96.

83. Yarotskyy V, Gao G, Peterson BZ, Elmslie KS. The Timothy syndrome mutation of cardiac CaV1.2 (L-type) channels: multiple altered gating mechanisms and pharmacological restoration of inactivation. J Physiol 2009;587:551-65.

84. Vatta M, Ackerman MJ, Ye B, Makielski JC, Ughanze EE, Taylor EW, et al. Mutant caveolin-3 induces persistent late sodium current and is associated with long-QT syndrome. Circulation 2006;114:2104-12.

85. Cronk LB, Ye B, Kaku T, Tester DJ, Vatta M, Makielski JC, et al. Novel mechanism for sudden infant death syndrome: persistent late sodium current secondary to mutations in caveolin-3. Heart Rhythm 2007;4:161-6.

86. Medeiros-Domingo A, Kaku T, Tester DJ, Iturralde-Torres P, Itty A, Ye B, et al. SCN4B-encoded sodium channel beta4 subunit in congenital long-QT syndrome. Circulation 2007;116:134-42.

87. Chen L, Marquardt ML, Tester DJ, Sampson KJ, Ackerman MJ, Kass RS. Mutation of an A-kinase-anchoring protein causes long-QT syndrome. Proc Natl Acad Sci U S A. 2007;104:20990-5.

88. Ueda K, Valdivia C, Medeiros-Domingo A, Tester DJ, Vatta M, Farrugia G, et al. Syntrophin mutation associated with long QT syndrome through activation of the nNOS-SCN5A macromolecular complex. Proc Natl Acad Sci U S A 2008;105:9355-60.

89. Splawski I, Timothy KW, Vincent GM, Atkinson DL, Keating MT. Molecular basis of the long-QT syndrome associated with deafness. N Engl J Med 1997;336:1562-7.

90. Neyroud N, Tesson F, Denjoy I, Leibovici M, Donger C, Barhanin J, et al. A novel mutation in the potassium channel gene KVLQT1 causes the Jervell and Lange-Nielsen cardioauditory syndrome. Nat Genet 1997;15:186-9.

91. Schulze-Bahr E, Wang Q, Wedekind H, Haverkamp W, Chen Q, Sun Y, et al. KCNE1 mutations cause jervell and Lange-Nielsen syndrome. Nat Genet 1997;17:267-8.

92. Schulze-Bahr E, Haverkamp W, Wedekind H, Rubie C, Hordt M, Borggrefe M, et al. Autosomal recessive long-QT syndrome (Jervell Lange-Nielsen syndrome) is genetically heterogeneous. Hum Genet 1997;100:573-6.

93. Tyson J, Tranebjaerg L, Bellman S, Wren C, Taylor JF, Bathen J, et al. IsK and KvLQT1: mutation in either of the two subunits of the slow component of the delayed rectifier potassium channel can cause Jervell and Lange-Nielsen syndrome. Hum Mol Genet 1997;6:2179-85.

94. Westenskow P, Splawski I, Timothy KW, Keating MT, Sanguinetti MC. Compound mutations: a common cause of severe long-QT syndrome. Circulation 2004;109:1834-41.

95. Yamaguchi M, Shimizu M, Ino H, Terai H, Hayashi K, Kaneda T, et al. Compound heterozygosity for mutations Asp611->Tyr in KCNQ1 and Asp609->Gly in KCNH2 associated with severe long QT syndrome. Clin Sci (Lond) 2005;108:143-50.

96. Kapplinger JD, Tester DJ, Salisbury BA, Carr JL, Harris-Kerr C, Pollevick GD, et al. Spectrum and prevalence of mutations from the first 2,500 consecutive unrelated patients referred for the FAMILION long QT syndrome genetic test. Heart Rhythm 2009;6:1297-303.

97. Zhang L, Vincent GM, Baralle M, Baralle FE, Anson BD, Benson DW, et all. An intronic mutation causes long QT syndrome. J Am Coll Cardiol 2004;44:1283-91.

98. Eddy CA, MacCormick JM, Chung SK, Crawford JR, Love DR, Rees MI, et al. Identification of large gene deletions and duplications in KCNQ1 and KCNH2 in patients with long QT syndrome. Heart Rhythm 2008;5:1275-81.

99. Tester DJ, Will ML, Haglund CM, Ackerman MJ. Effect of clinical phenotype on yield of long QT syndrome genetic testing. J Am Coll Cardiol 2006;47:764-8.

100. Donger C, Denjoy I, Berthet M, Neyroud N, Cruaud C, Bennaceur M, et al. KVLQT1 C-terminal missense mutation causes a forme fruste long-QT syndrome. Circulation 1997;96:2778-81.

101. Shimizu W, Horie M, Ohno S, Takenaka K, Yamaguchi M, Shimizu M, et al. Mutation site-specific differences in arrhythmic risk and sensitivity to sympathetic stimulation in the LQT1 form of congenital long QT syndrome: multicenter study in Japan. J Am Coll Cardiol 2004;44:117-25.

102. Shimizu W, Moss AJ, Wilde AA, Towbin JA, Ackerman MJ, January CT, et al. Genotype-phenotype aspects of type 2 long QT syndrome. J Am Coll Cardiol 2009;54:2052-62.

103. Liu JF, Moss AJ, Jons C, Benhorin J, Schwartz PJ, Spazzolini C, et al. Mutation-specific risk in two genetic forms of type 3 long QT syndrome. Am J Cardiol 2010;105:210-3.

104. Bankston JR, Yue M, Chung W, Spyres M, Pass RH, Silver E, et al. A novel and lethal de novo LQT-3 mutation in a newborn with distinct molecular pharmacology and therapeutic response. PLoS One 2007;2:e1258.

105. Ackerman MJ, Tester DJ, Porter CJ, Edwards WD. Molecular diagnosis of the inherited long-QT syndrome in a woman who died after near-drowning. N Engl J Med 1999;341:1121-5.

106. Ali RH, Zareba W, Moss AJ, Schwartz PJ, Benhorin J, Vincent GM, et al. Clinical and genetic variables associated with acute arousal and nonarousal-related cardiac events among subjects with long QT syndrome. Am J Cardiol 2000;85:457-61.

107. Moss AJ, Zareba W, Benhorin J, Locati EH, Hall WJ, Robinson JL, et al. ECG T-wave patterns in genetically distinct forms of the hereditary long QT syndrome. Circulation 1995;92:2929-34.

108. Zhang L, Timothy KW, Vincent GM, Lehmann MH, Fox J, Giuli LC, et al. Spectrum of ST-T-wave patterns and repolarization parameters in congenital long-QT syndrome: ECG findings identify genotypes. Circulation 2000;102:2849-55.

109. Shimizu W, Antzelevitch C. Cellular basis for long QT, transmural dispersion of repolarization, and torsade de pointes in the long QT syndrome. J Electrocardiol 1999;32:177-84.

110. Shimizu W, Antzelevitch C. Differential response of transmural dispersion of repolarization and torsade de pointes to beta-adrenergic agonists and antagonists in three models of the long QT syndrome. J Electrocardiol 1999;32:150.

111. Noda T, Shimizu W, Satomi K, Suyama K, Kurita T, Aihara N, et al. Classification and mechanism of Torsade de Pointes initiation in patients with congenital long QT syndrome. Eur Heart J 2004;25:2149-54.

112. Tan HL, Bardai A, Shimizu W, Moss AJ, Schulze-Bahr E, Noda T, et al. Genotype-specific onset of arrhythmias in congenital long-QT syndrome: possible therapy implications. Circulation 2006;114:2096-103.
113. Zareba W, Moss AJ, Schwartz PJ, Vincent GM, Robinson JL, Priori SG, et al. Influence of genotype on the clinical course of the long-QT syndrome. International Long-QT Syndrome Registry Research Group. N Engl J Med 1998;339:960-5.

114. Locati EH, Zareba W, Moss AJ, Schwartz PJ, Vincent GM, Lehmann MH, et al. Age- and sex-related differences in clinical manifestations in patients with congenital long-QT syndrome: findings from the International LQTS Registry. Circulation 1998;97:2237-44.

115. Goldenberg I, Moss AJ. Long QT syndrome. J Am Coll Cardiol 2008;51:2291-300.

116. Spazzolini C, Mullally J, Moss AJ, Schwartz PJ, McNitt S, Ouellet G, et al. Clinical implications for patients with long QT syndrome who experience a cardiac event during infancy. J Am Coll Cardiol 2009;54:832-7.

117. Goldenberg I, Moss AJ, Peterson DR, McNitt S, Zareba W, Andrews ML, et al. Risk factors for aborted cardiac arrest and sudden cardiac death in children with the congenital long-QT syndrome. Circulation 2008;117:2184-91.

118. Hobbs JB, Peterson DR, Moss AJ, McNitt S, Zareba W, Goldenberg I, et al. Risk of aborted cardiac arrest or sudden cardiac death during adolescence in the long-QT syndrome. JAMA 2006;296:1249-54.

119. Seth R, Moss AJ, McNitt S, Zareba W, Andrews ML, Qi M, et al. Long QT syndrome and pregnancy. J Am Coll Cardiol 2007;49:1092-8.
120. Priori SG, Schwartz PJ, Napolitano C, Bloise R, Ronchetti E, Grillo M, et al. Risk stratification in the long-QT syndrome. N Engl J Med 2003;348:1866-74.

121. Goldenberg I, Moss AJ, Bradley J, Polonsky S, Peterson DR, McNitt S, et al. Long-QT syndrome after age 40. Circulation 2008;117:2192-201.

122. Kimbrough J, Moss AJ, Zareba W, Robinson JL, Hall WJ, Benhorin J, et al. Clinical implications for affected parents and siblings of probands with long-QT syndrome. Circulation 2001;104:557-62.

123. Adamantidis MM. [Mechanisms of action of class III antiarrhythmia agents]. Arch Mal Coeur Vaiss 1995;88:33-40.

124. Gury C, Canceil O, Iaria P. [Antipsychotic drugs and cardiovascular safety: current studies of prolonged QT interval and risk of ventricular arrhythmia]. Encephale 2000;26:62-72.

125. Lehmann MH, Hardy S, Archibald D, quart B, MacNeil DJ. Sex difference in risk of torsade de pointes with d,l-sotalol. Circulation 1996;94:2535-41.

126. Drici MD, Burklow TR, Haridasse V, Glazer RI, Woosley RL. Sex hormones prolong the QT interval and downregulate potassium channel expression in the rabbit heart. Circulation 1996;94:1471-4.
127. Sanguinetti MC, Chen J, Fernandez D, Kamiya K, Mitcheson J, Sanchez-Chapula JA. Physicochemical basis for binding and voltagedependent block of hERG channels by structurally diverse drugs. Novartis Found Symp 2005;266:159-66.

128. Takemasa H, Nagatomo T, Abe H, Kawakami K, Igarashi T, Tsurugi T, et al. Coexistence of hERG current block and disruption of protein trafficking in ketoconazole-induced long QT syndrome. Br J Pharmacol 2008;153:439-47.

129. Wehrens XH. Structural determinants of potassium channel blockade and drug-induced arrhythmias. Handb Exp Pharmacol 2006;171:123-57.

130. Sanguinetti MC, Mitcheson JS. Predicting drug-hERG channel interactions that cause acquired long QT syndrome. Trends Pharmacol Sci 2005;26:119-24.

131. Sanguinetti MC, Tristani-Firouzi M. hERG potassium channels and cardiac arrhythmia. Nature 2006;440:463-9.

132. Dennis A, Wang L, Wan X, Ficker E. hERG channel trafficking: novel targets in drug-induced long QT syndrome. Biochem Soc Trans 2007;35:1060-3.

133. De Ponti F, Poluzzi E, Cavalli A, Recanatini M, Montanaro N. Safety of non-antiarrhythmic drugs that prolong the QT interval or induce torsade de pointes: an overview. Drug Saf 2002;25:263-86.

134. Chvilicek JP, Hurlbert BJ, Hill GE. Diuretic-induced hypokalaemia inducing torsades de pointes. Can J Anaesth 1995;42:1137-9.

135. Guo J, Massaeli H, Xu J, Jia Z, Wigle JT, Mesaeli N, et al. Extracellular K+ concentration controls cell surface density of IKr in rabbit hearts and of the HERG channel in human cell lines. J Clin Invest 2009;119:2745-57.

136. Mullins FM, Stepanovic SZ, Desai RR, George AL Jr, Balser JR. Extracellular sodium interacts with the HERG channel at an outer pore site. J Gen Physiol 2002;120:517-37.

137. January CT, Riddle JM, Salata JJ. A model for early afterdepolarizations: induction with the Ca2+ channel agonist Bay K 8644. Circ Res 1988;62:563-71.

138. Cohen IS, DiFrancesco D, Mulrine NK, Pennefather P. Internal and external K+ help gate the inward rectifier. Biophys J 1989;55:197-202.

139. Morita H, Zipes DP, Morita ST, Wu J. Mechanism of U wave and polymorphic ventricular tachycardia in a canine tissue model of Andersen-Tawil syndrome. Cardiovasc Res 2007;75:510-8.

140. Surawicz B, Lepeschkin E. The electrocardiographic pattern of hypopotassemia with and without hypocalcemia. Circulation 1953;8:801-28.

141. Leclercq JF, Coumel P, Maison-Blanche P, Cauchemez B, Zimmermann M, Chouty F, et al. [Mechanisms determining sudden death. A cooperative study of 69 cases recorded using the Holter method]. Arch Mal Coeur Vaiss 1986;79:1024-33.

142. Kubota T, Shimizu W, Kamakura S, Horie M. Hypokalemiainduced long QT syndrome with an underlying novel missense mutation in S4-S5 linker of KCNQ1. J Cardiovasc Electrophysiol 2000;11:1048-54.

143. Gums JG. Magnesium in cardiovascular and other disorders. Am J Health Syst Pharm 2004;61:1569-76.

144. Dalmas O. Magnesium selective ion channels. Biophys J 2007;93:3729-30.

145. Kulkarni P, Bhattacharya S, Petros AJ. Torsade de pointes and long QT syndrome following major blood transfusion. Anaesthesia 1992;47:125-7.

146. Gonzalez MM, Cavalcanti TC, Vianna CB, Timerman S. Hypomagnesaemia causing QT interval prolongation and torsade de pointes in an alcoholic patient. Resuscitation 2006;70:346-7.

147. Nguyen H, Zaroff JG. Neurogenic stunned myocardium. Curr Neurol Neurosci Rep 2009;9:486-491.

148. Kono T, Morita H, Kuroiwa T, Onaka H, Takatsuka H, Fujiwara A. Left ventricular wall motion abnormalities in patients with subarachnoid hemorrhage: neurogenic stunned myocardium. J Am Coll Cardiol 1994;24:636-40.

149. Jarquin-Valdivia AA, Rich AT, Yarbrough JL, Thompson RC. Intraventricular colloid cyst, hydrocephalus and neurogenic stunned myocardium. Clin Neurol Neurosurg 2005;107:361-5.

150. Wittstein IS, Thiemann DR, Lima JA, Baughman KL, Schulman SP, Gerstenblith G, et al. Neurohumoral features of myocardial stunning due to sudden emotional stress. N Engl J Med 2005;352:539-48.

151. Bulsara KR, McGirt MJ, Liao L, Villavicencio AT, Borel C, Alexander MJ, et al. Use of the peak troponin value to differentiate myocardial infarction from reversible neurogenic left ventricular dysfunction associated with aneurysmal subarachnoid hemorrhage. J Neurosurg 2003;98:524-8.

152. Su Z, Limberis J, Martin RL, Xu R, Kolbe K, Heinemann SH, et al. Functional consequences of methionine oxidation of hERG potassium channels. Biochem Pharmacol 2007;74:702-11.

153. Weissenburger J, Davy JM, Chezalviel F, Ertzbischoff O, Poirier JM, Engel F, et al. Arrhythmogenic activities of antiarrhythmic drugs in conscious hypokalemic dogs with atrioventricular block: comparison between quinidine, lidocaine, flecainide, propranolol and sotalol. J Pharmacol Exp Ther 1991;259:871-83.

154. Peters RH, Wever EF, Hauer RN, Wittkampf FH, Robles de Medina EO. Bradycardia dependent QT prolongation and ventricular fibrillation following catheter ablation of the atrioventricular junction with radiofrequency energy. Pacing Clin Electrophysiol 1994;17:108-12.

155. Hreiche R, Morissette P, Turgeon J. Drug-induced long QT syndrome in women: review of current evidence and remaining gaps. Gend Med 2008;5:124-35.

156. Rodriguez I, Kilborn MJ, Liu XK, Pezzullo JC, Woosley RL. Druginduced QT prolongation in women during the menstrual cycle. JAMA 2001;285:1322-6.

157. Vatta M, Towbin JA. Mutations in KCNE1 in long QT syndrome (LQTS): insights into mechanism of LQTS and drug sensitivity? Heart Rhythm 2006;3:1041-2.

158. Ohno S, Toyoda F, Zankov DP, Yoshida H, Makiyama T, Tsuji K, et al. Novel KCNE3 mutation reduces repolarizing potassium current and associated with long QT syndrome. Hum Mutat 2009;30:557-63.
159. Roden DM, Viswanathan PC. Genetics of acquired long QT syndrome. J Clin Invest 2005;115:2025-32.

160. Laitinen P, Fodstad H, Piippo K, Swan H, Toivonen L, Viitasalo M, et al. Survey of the coding region of the HERG gene in long QT syndrome reveals six novel mutations and an amino acid polymorphism with possible phenotypic effects. Hum Mutat 2000;15:580-1.

161. Yang P, Kanki H, Drolet B, Yang T, Wei J, Viswanathan PC, et al. Allelic variants in long-QT disease genes in patients with drugassociated torsades de pointes. Circulation 2002;105:1943-8.

162. Ackerman MJ, Tester DJ, Jones GS, Will ML, Burrow CR, Curran ME. Ethnic differences in cardiac potassium channel variants: implications for genetic susceptibility to sudden cardiac death and genetic testing for congenital long QT syndrome. Mayo Clin Proc 2003;78:1479-87.

163. Anson BD, Ackerman MJ, Tester DJ, Will ML, Delisle BP, Anderson CL, et al. Molecular and functional characterization of common polymorphisms in HERG (KCNH2) potassium channels. Am J Physiol Heart Circ Physiol 2004;286:H2434-41. **164.** Roden DM. Long QT syndrome: reduced repolarization reserve and the genetic link. J Intern Med 2006;259:59-69.

165. Splawski I, Timothy KW, Tateyama M, Clancy CE, Malhotra A, Beggs AH, et al. Variant of SCN5A sodium channel implicated in risk of cardiac arrhythmia. Science 2002;297:1333-6.

166. Ackerman MJ, Splawski I, Makielski JC, Tester DJ, Will ML, Timothy KW, et al. Spectrum and prevalence of cardiac sodium channel variants among black, white, Asian, and Hispanic individuals: implications for arrhythmogenic susceptibility and Brugada/long QT syndrome genetic testing. Heart Rhythm 2004;1:600-7.

167. Makita N, Horie M, Nakamura T, Ai T, Sasaki K, Yokoi H, et al. Drug-induced long-QT syndrome associated with a subclinical SCN5A mutation. Circulation 2002;106:1269-74.

168. Kubota T, Horie M, Takano M, Yoshida H, Takenaka K, Watanabe E, et al. Evidence for a single nucleotide polymorphism in the KCNQ1 potassium channel that underlies susceptibility to life-threatening arrhythmias. J Cardiovasc Electrophysiol 2001;12:1223-9.

169. Sicouri S, Antzelevitch C. A subpopulation of cells with unique electrophysiological properties in the deep subepicardium of the canine ventricle. The M Cell Circ Res 1991;68:1729-41.

170. Antzelevitch C, Sicouri S, Litovsky SH, Lukas A, Krishnan SC, Di Diego JM, et al. Heterogeneity within the ventricular wall. Electrophysiology and pharmacology of epicardial, endocardial, and M cells. Circ Res 1991;69:1427-49.

171. Sicouri S, Antzelevitch C. Afterdepolarizations and triggered activity develop in a select population of cells (M cells) in canine ventricular myocardium: the effects of acetylstrophanthidin and Bay K 8644. Pacing Clin Electrophysiol 1991;14:1714-20.

172. Antzelevitch C, Belardinelli L. The role of sodium channel current in modulating transmural dispersion of repolarization and arrhythmogenesis. J Cardiovasc Electrophysiol 2006;17:S79-S85.

173. Shimizu W, Antzelevitch C. Cellular basis for the ECG features of the LQT1 form of the long-QT syndrome: effects of beta-adrenergic agonists and antagonists and sodium channel blockers on transmural dispersion of repolarization and torsade de pointes. Circulation 1998;98:2314-22.

174. Shimizu W, Antzelevitch C. Sodium channel block with mexiletine is effective in reducing dispersion of repolarization and preventing torsade des pointes in LQT2 and LQT3 models of the long-QT syndrome. Circulation 1997;96:2038-47.

175. Shimizu W, Antzelevitch C. Differential effects of beta-adrenergic agonists and antagonists in LQT1, LQT2 and LQT3 models of the long QT syndrome. J Am Coll Cardiol 2000;35:778-86.

176. Strickberger SA, Benson DW, Biaggioni I, Callans DJ, Cohen MI, Ellenbogen KA, et al; American Heart Association Councils on Clinical Cardiology, Cardiovascular Nursing, Cardiovascular Disease in the Young, and Stroke; Quality of Care and Outcomes Research Interdisciplinary Working Group; American College of Cardiology Foundation; Heart Rhythm Society; American Autonomic Society. AHA/ACCF Scientific Statement on the evaluation of syncope: from the American Heart Association Councils on Clinical Cardiology, Cardiovascular Nursing, Cardiovascular Disease in the Young, and Stroke, and the Quality of Care and Outcomes Research Interdisciplinary Working Group; and the American College of Cardiology Foundation: in collaboration with the Heart Rhythm Society: endorsed by the American Autonomic Society. Circulation 2006;113:316-27.

177. Johnson JN, Hofman N, Haglund CM, Cascino GD, Wilde AA, Ackerman MJ. Identification of a possible pathogenic link between congenital long QT syndrome and epilepsy. Neurology 2009;72:224-31.
178. Lepeschkin E, Surawicz B. The measurement of the Q-T interval of the electrocardiogram. Circulation 1952;6:378-88.

179. Rautaharju PM, Surawicz B, Gettes LS, Bailey JJ, Childers R, Deal BJ, et al; American Heart Association Electrocardiography and Arrhythmias Committee, Council on Clinical Cardiology; American College of Cardiology Foundation; Heart Rhythm Society. AHA/ACCF/HRS recommendations for the standardization and interpretation of

the electrocardiogram: part IV: the ST segment, T and U waves, and the QT interval: a scientific statement from the American Heart Association Electrocardiography and Arrhythmias Committee, Council on Clinical Cardiology; the American College of Cardiology Foundation; and the Heart Rhythm Society. Endorsed by the International Society for Computerized Electrocardiology. J Am Coll Cardiol 2009;53:982-91.

180. Lepeschkin E, Surawicz B. The duration of the Q-U interval and its components in electrocardiograms of normal persons. Am Heart J 1953;46:9-20.

181. Surawicz B, Parikh SR. Differences between ventricular repolarization in men and women: description, mechanism and implications. Ann Noninvasive Electrocardiol 2003;8:333-40.

182. Chen S, Zhang L, Bryant RM, Vincent GM, Flippin M, Lee JC, et al. KCNQ1 mutations in patients with a family history of lethal cardiac arrhythmias and sudden death. Clin Genet 2003;63:273-82.
183. Li C, Hu D, Qin X, Li Y, Li P, Liu W, et al. Clinical features and management of congenital long QT syndrome: a report on 54 patients from a national registry. Heart Vessels 2004;19:38-42.

184. Goldenberg I, Mathew J, Moss AJ, McNitt S, Peterson DR, Zareba W, et al. Corrected QT variability in serial electrocardiograms in long QT syndrome: the importance of the maximum corrected QT for risk stratification. J Am Coll Cardiol 2006;48:1047-52.

185. Swan H, Toivonen L, Viitasalo M. Rate adaptation of QT intervals during and after exercise in children with congenital long QT syndrome. Eur Heart J 1998;19:508-13.

186. Takenaka K, Ai T, Shimizu W, Kobori A, Ninomiya T, Otani H, et al. Exercise stress test amplifies genotype-phenotype correlation in the LQT1 and LQT2 forms of the long-QT syndrome. Circulation 2003;107:838-44.

187. Schwartz PJ, Priori SG, Locati EH, Napolitano C, Cantu F, Towbin JA, et al. Long QT syndrome patients with mutations of the SCN5A and HERG genes have differential responses to Na+ channel blockade and to increases in heart rate. Implications for gene-specific therapy. Circulation 1995;92:3381-6.

188. Ackerman MJ, Khositseth A, Tester DJ, Hejlik JB, Shen WK, Porter CB. Epinephrine-induced QT interval prolongation: a genespecific paradoxical response in congenital long QT syndrome. Mayo Clin Proc 2002;77:413-21.

189. Noda T, Takaki H, Kurita T, Suyama K, Nagaya N, Taguchi A, et al. Gene-specific response of dynamic ventricular repolarization to sympathetic stimulation in LQT1, LQT2 and LQT3 forms of congenital long QT syndrome. Eur Heart J 2002;23:975-83.

190. Tester DJ, Will ML, Ackerman MJ. Mutation detection in congenital long QT syndrome: cardiac channel gene screen using PCR, dHPLC, and direct DNA sequencing. Methods Mol Med 2006; 128:181-207.

191. Zipes DP, Camm AJ, Borggrefe M, Buxton AE, Chaitman B, Fromer M, et al; American College of Cardiology/American Heart Association Task Force; European Society of Cardiology Committee for Practice Guidelines; European Heart Rhythm Association; Heart Rhythm Society. ACC/AHA/ESC 2006 Guidelines for Management of Patients With Ventricular Arrhythmias and the Prevention of Sudden Cardiac Death: a report of the American College of Cardiology/American Heart Association Task Force and the European Society of Cardiology Committee for Practice Guidelines (writing committee to develop Guidelines for Management of Patients With Ventricular Arrhythmias and the Prevention of Sudden Cardiac Death): developed in collaboration with the European Heart Rhythm Association and the Heart Rhythm Society. Circulation 2006;114:e385-484.

192. Aomine M, Tatsukawa Y, Ehara T. [Antiarrhythmic effects of magnesium on single ventricular myocytes]. Kokyu To Junkan 1992;40:677-83.

193. Bristow M. Antiadrenergic therapy of chronic heart failure: surprises and new opportunities. Circulation 2003;107:1100-2.

194. Crotti L, Spazzolini C, Schwartz PJ, Shimizu W, Denjoy I, Schulze-Bahr E, et al. The common long-QT syndrome mutation

KCNQ1/A341V causes unusually severe clinical manifestations in patients with different ethnic backgrounds: toward a mutation-specific risk stratification. Circulation 2007;116:2366-75.

195. Kobori A, Sarai N, Shimizu W, Nakamura Y, Murakami Y, Makiyama T, et al. Additional gene variants reduce effectiveness of beta-blockers in the LQT1 form of long QT syndrome. J Cardiovasc Electrophysiol 2004;15:190-9.

196. Liggett SB, Mialet-Perez J, Thaneemit-Chen S, Weber SA, Greene SM, Hodne D, et al. A polymorphism within a conserved beta(1)-adrenergic receptor motif alters cardiac function and beta-blocker response in human heart failure. Proc Natl Acad Sci U S A 2006;103:11288-93.

197. Bristow MR, Murphy GA, Krause-Steinrauf H, Anderson JL, Carlquist JF, Thaneemit-Chen S, et al. An alpha2C-adrenergic receptor polymorphism alters the norepinephrine-lowering effects and therapeutic response of the beta-blocker bucindolol in chronic heart failure. Circ Heart Fail 2010;3:21-8.

198. Compton SJ, Lux RL, Ramsey MR, Strelich KR, Sanguinetti MC, Green LS, et al. Genetically defined therapy of inherited long-QT syndrome. Correction of abnormal repolarization by potassium. Circulation 1996;94:1018-22.

199. Etheridge SP, Compton SJ, Tristani-Firouzi M, Mason JW. A new oral therapy for long QT syndrome: long-term oral potassium

improves repolarization in patients with HERG mutations. J Am Coll Cardiol 2003;42:1777-82.

200. Moss AJ, McDonald J. Unilateral cervicothoracic sympathetic ganglionectomy for the treatment of long QT interval syndrome. N Engl J Med 1971;285:903-4.

201. Schwartz PJ, Locati EH, Moss AJ, Crampton RS, Trazzi R, Ruberti U. Left cardiac sympathetic denervation in the therapy of congenital long QT syndrome. A worldwide report. Circulation 1991;84:503-11.

202. Schwartz PJ, Priori SG, Cerrone M, Spazzolini C, Odero A, Napolitano C, et al. Left cardiac sympathetic denervation in the management of high-risk patients affected by the long-QT syndrome. Circulation 2004;109:1826-33.

203. Li J, Wang L, Wang J. Video-assisted thoracoscopic sympathectomy for congenital long QT syndromes. Pacing Clin Electrophysiol 2003;26:870-3.

204. Li J, Liu Y, Yang F, Jiang G, Li C, Hu D, et al. Video-assisted thoracoscopic left cardiac sympathetic denervation: a reliable minimally invasive approach for congenital long-QT syndrome. Ann Thorac Surg 2008;86:1955-8.

205. Collura CA, Johnson JN, Moir C, Ackerman MJ. Left cardiac sympathetic denervation for the treatment of long QT syndrome and catecholaminergic polymorphic ventricular tachycardia using video-assisted thoracic surgery. Heart Rhythm 2009;6:752-9.